Status of knowledge of the broad-nosed weevils of Colombia (Coleoptera, Curculionidae, Entiminae)

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Abstract
Broad-nosed weevils in the subfamily Entiminae (Coleoptera: Curculionidae) are highly diverse, not only in terms of number of species, but also in their sizes, forms and colours. There are eight tribes, 50 genera and 224 entimine species recorded from Colombia: seven genera and 142 species are considered endemic and only a handful of species, which are recognised as pests of Citrus or potatoes, are broadly known. The large diversity of this subfamily in the country is only superficially known and even though genus level identifications are generally achievable, species identification remains quite challenging, due in part to limited access to broadly-scattered basic information. Summaries of available information and bibliographic resources for each of the tribes represented in Colombia are offered, along with a checklist of the species of Entiminae recorded from the country, obtained from literature and a pictorial key for tribal recognition. New combinations are proposed for eight species of the genus Lanterius Alonso-Zarazaga & Lyal. Information on the distribution of entimine species in Colombia is compiled for the first time, including complete references to each original description and available taxonomic revisions. About a third of the species of Entiminae remain as recorded from the country without specific locality information. In addition, genus level distributional maps are presented, generated from data obtained from four Colombian entomological collections. Lastly, some challenges for entimine identification in Colombia, which likely extend throughout the Neotropical region, are briefly discussed. This contribution aims, in part, to facilitate and promote entimine research in northern South America.
Resumen
Los gorgojos de pico corto en la subfamilia Entiminae (Coleoptera: Curculionidae) son altamente diversos, no sólo en términos del número de especies, sino además en tamaños, formas y colores. Existen ocho tribus, 50 géneros y 224 especies entiminos registradas en Colombia: siete géneros y 142 especies son consideradas endémicas, y sólo un puñado, las cuales son reconocidas como plagas de citricos o papa, son ampliamente conocidas. La gran diversidad de esta subfamilia en el país sólo se conoce superficialmente y aunque identificar géneros es generalmente alcanzable, la identificación de especies sigue siendo un desafío, debido en parte al acceso limitado a información básica ampliamente dispersa. Se ofrecen resúmenes de información disponible y recursos bibliográficos para cada una de las tribus representadas en Colombia, junto con la lista de especies de Entiminae registradas para el país, obtenida a partir de la literatura, junto con una clave pictórica para el reconocimiento de tribus. Se proponen nuevas combinaciones para ocho especies del género Lanterius Alonso-Zarazaga & Lyal. Información sobre la distribución de las especies de Entiminae en Colombia se recopila por primera vez, incluyendo referencias completas a cada una de las descripciones originales y a las revisiones taxonómicas disponibles. Cerca de un tercio de las especies de Entiminae permanecen registradas para el país sin información específica de localidad. Adicionalmente, se presentan mapas de distribución a nivel de género, producidos a partir de datos obtenidos de cuatro colecciones entomológicas colombianas. Por último, algunas de las dificultades para la identificación de entiminos en Colombia, que probablemente se extienden a lo largo de la Región Neotropical, son brevemente discutidas. Esta contribución busca en parte facilitar y promover la investigación en Entiminae en el norte de Suramérica.

Keywords
Distribution, faunistics, literature, Neotropics, original description, South America, taxonomy

Palabras clave
Distribución, faunística, literatura, neotrópico, descripción original, Suramérica, taxonomía

Introduction
Weevils in the subfamily Entiminae Schönherr, 1823 (Schönherr 1823, col. 1138), commonly known as broad-nosed weevils (or “vaquitas” or “mulitas” in Spanish), constitute one of the most diverse, abundant and widespread groups of weevils worldwide, including well over 12000 described species (Oberprieler et al. 2007). The subfamily includes some of the most recognisable, charismatic and stunning weevil forms; for example, the genus Eupholus Boisduval, 1835 (Boisduval 1835, 363), from New Guinea, commonly known as “smurf-weevils” (see Van Dam et al. 2017), the genus Pachyrhynchus Germar, 1824 (Germar 1824, 336), in Southeast Asia (e.g. Háva and Rukmane 2018), sometimes called polka-dot weevils and the genera Briarius Fischer de Waldheim, 1829 (Fischer de Waldheim 1829, 88); formerly Lamprocyphus Marshall, 1922 (Marshall 1922b, 184; see Lanteri and del Río 2003) and Entimus Germar, 1817 (Germar 1817, 341), both from South America.

In taxonomic terms, the subfamily Entiminae has been recognised as such since Thompson (1992) and has been divided into as few as five tribes (Marvaldi 1997,
1998) to as many as 55 tribes (Alonso-Zarazaga and Lyal 1999); 54 according to Bouchard et al. (2011) is the most widely-accepted number of tribes. A summary of the taxonomic history of the Entiminae is provided by Marvaldi et al. (2014).

**Biological generalities of entimines**

Amongst curculionids, entimines are usually easy to recognise by the typical short and broad form of their rostrum, which is accompanied, in most cases, by conspicuous mandibular scars left by the breaking off of a deciduous process of the mandible at the time when the adult emerges from the pupal stage (Anderson and Howden 2002; see Fig. 1A). Entimines are phytophagous, trending to polyphagous and oligophagous; the larvae feed underground on the roots of plants, whereas the adults feed on the leaves (Marvaldi et al. 2014). Relatively few species of entimines (considering the whole diversity of the subfamily) are recognised as pests of economical importance; usually, a species only becomes a problem when populations are large. Especially relevant for Colombia are *Compsus viridivittatus* (Guérin-Méneville, 1855) (Guérin-Méneville 1855, 592), a *Citrus* pest in the tribe Eustylini (Cano et al. 2002a, b; Gallego et al. 2012, O’Brien and Peña 2012) and the ‘Potato shooter’ *Leschenius vulcanorum* (Kirsch, 1889) (Kirsch 1889, 17) in the tribe Naupactini (Canchala 1992; Peña 2001; Cortázar Gómez et al. 2012).

A thorough summary of the natural history and morphology (of both adults and larvae) of the Entiminae is provided by Marvaldi et al. (2014). Additional information regarding morphological features of the adults and the larvae can be found in Marvaldi and Lanteri (2005) and Marvaldi (1997, 1998), respectively.

Most tribes are limited to particular biogeographic regions of the world (Marvaldi et al. 2014). Their distributions, in general, are more a reflection of habitat rather than host plant preferences (Anderson 1993, 2018; Marvaldi et al. 2014). The wide range of plants they feed on, paired with the strong association of species to particular habitats and geographic regions (e.g. Lanteri 1992; Anderson and Lanteri 2000; Franz and Girón 2009; Franz 2010b; Girón and Franz 2010; Franz 2011; Mazo-Vargas 2011, del Río and Lanteri 2011a; Girón and Franz 2012, del Río et al. 2015), makes entimines a potential model group for biogeographic studies and the identification of areas of endemism. There is also evidence of altitudinal stratification of species (Girón and Franz 2010).

As a consequence of their diversity and abundance, entimines are commonly found in the field, including urbanised areas and are, therefore, frequently found in large numbers in biological collections (see Girón and Cardona-Duque 2018). There is also a trend to find large numbers of individuals of single economical-ly-important species in cultivars, whereas only few specimens of several different species can be found associated with forested or less disturbed areas (pers. obs.). Furthermore, it is common that specimens in collections are only identified to family or, at best, sorted to subfamily, as is often the case in Colombia (see Girón and Cardona-Duque 2018).
Figure 1. Select morphological features of Entiminae: A, G ventral view of rostrum: A right deciduous process still attached, left mandible with mandibular scar, maxillae fully covered by prementum (Naupactini) G mandibular scars well developed, maxillae partially covered by prementum (Anypotactus sp.) B, C frontal view of head indicating position of epistoma, nasal plate and mandibular scars: B Naupactini, with median furrow C Eustylini, with median fovea D–F lateral view of head and anterior section of prothorax: D Tanymecini, with anterior margin of prothorax straight and postocular setae clustered as a tuft E Eustylini, with anterior margin of prothorax slightly sinuate and without postocular setae F Lordopini, with anterior margin of prothorax strongly sinuate as to form well-developed postocular lobe, with postocular setae forming a fringe H–K profemora in frontal view (top end of drawings articulate to coxa, bottom end articulate to tibia): H Compsus sp., regularly shaped profemur I Eustylus sp., toothed profemur J Hadromeropsis sp. enlarged profemur K Anypotactus sp., clavate and toothed profemur L, M apical region of left metatibia (left margin on drawing is anterior (inner) in the beetle; right margin on drawing is posterior (outer) in the beetle), black triangles indicate posterior corner of metatibia: L Eustylini, posterior corner of metatibia rounded M Naupactini, posterior corner of metatibia angulate N–P anterior section of elytra in dorsal view: N shoulders absent O shoulders oblique, weakly developed P shoulders well developed.
Identification challenges

One of the most relevant references for Neotropical Broad-nosed weevils is the ‘Biología Centrali-Americana’ by Sharp and Champion (1911), although it includes only scarce information on Colombian taxa. Even though many genera have been revised (see below under the treatment for each tribe), species identification of Neotropical and, especially northern South American, entimines remains highly challenging (e.g. Girón 2006). For Colombia, in particular, the fact that most type specimens are deposited in collections overseas, combined with the lack of identified material in national collections and compounded by a lack of specialists in the region, makes species identification a very difficult task at this time. In addition, one of the main difficulties for identifying entimine taxa is the limited access to information, which has been changing in the digital era. Many publications, including original descriptions and revisions are currently available online, some of them for free through the Biodiversity Heritage Library (https://www.biodiversitylibrary.org/). Nevertheless, some key publications with full references linking to those original descriptions (e.g. Wibmer and O’Brien 1986a; Alonso-Zarazaga and Lyal 1999) remain available only on paper and are essentially inaccessible to the broad community, especially in Latin American countries. An effort has been made in this paper to incorporate references and links to as many relevant publications as possible.

This contribution compiles and summarises the available information for the Entiminae recorded from Colombia. A general pictorial key to diagnose tribes is given. For each tribe represented in the country, a list of morphological characters for tribal recognition is offered, along with distributional information and bibliographic resources. A list of species is provided, including bibliographic records of distributional data within the country, where available. In addition, genus-level information has been recorded from specimens identified in Colombian national entomological collections, further providing information about distributions of entimines in the country.

Materials and methods

Species list

The list of species of Entiminae recorded from Colombia, as well as general species distributions, were extracted from the annotated checklist of the weevils of South America by Wibmer and O’Brien (1986a), the list of species of Entiminae by Morrone (1999), in addition to more recent revisions or new descriptions, where available. Tribal concepts and general distributions of genera are in accordance with Alonso-Zarazaga and Lyal (1999). The full list of species is included here as Suppl. material 1 and available as a checklist via GBIF (Girón 2020, https://doi.org/10.15472/jdwfao); this online resource will be updated whenever new species are added to the Colombian fauna of Entiminae.
Only presence in Colombia was obtained from the main references (Wibmer and O’Brien 1986a; Morrone 1999 and Alonso-Zarazaga and Lyal 1999); therefore, original descriptions for each species were checked to obtain distributional records within the country. Localities taken from the 1800s’ to mid-1900s’ publications might not reflect current departments or municipalities (e.g. ‘Bogotá’ as a locality in 1857 does not strictly correspond to current ‘Cundinamarca: Bogotá’). In those cases, locality is quoted as presented in the original description. Department was added for unambiguous localities (e.g. Antioquia was added to original localities recorded as ‘Medellín’; Cundinamarca added for Fusagasugá; Meta added for Villavicencio, Boyacá added for Muzo). Localities in square brackets ‘[]’ in the list of species indicate locality spelling in the original description. Type locality is recorded as ‘Bogotá’ for species described by Kirsch (1868) given the title of the paper: “Beiträge zur Käferfauna von Bogotá”. A question mark ‘?’ indicates that the locality record is doubtful and needs confirmation. Specific localities from countries other than Colombia are omitted here. For some of Schönherr’s taxa, the page number corresponds to a column number in the publication and is indicated by ‘col.’ (e.g. Entiminae Schönherr 1823, col. 1138).

Material examined

Specimens from entomological collections in Colombia were revised and distributional data at the generic level were recorded (Suppl. material 2). The insect collection abbreviations are adopted, for the most part, from Evenhuis (2020) as follows:

IAvH  Instituto Alexander von Humboldt, Villa de Leyva, Colombia, https://doi.org/10.15472/vmpedy [databased July 2013]
MEPB  Museo Entomológico de Piedras Blancas, Comfenalco Antioquia, Colombia, https://doi.org/10.15472/xblw2v [databased March 2013]
MPUJ  Museo Javeriano de Historia Natural Lorenzo Uribe, S.J., Bogotá, Colombia, https://doi.org/10.15472/xmukx8 [partly databased October 2015]

A few additional records were obtained from ASUCOB (Arizona State University Charles W. O’Brien Collection, Arizona, USA https://doi.org/10.15468/eeqtxt0). Information obtained from collections is summarised for each tribe and identified genus.

Genus-level identifications were obtained using available keys (van Emden 1944a, b) and diagnoses (Sharp and Champion 1911) and confirmed by consulting experts in Curculionidae (Dr. Robert Anderson, Canadian Museum of Natural History, Ottawa, Canada and Dr. Charles O’Brien, USA), who have access to reference collections. The recognition section presented here for each tribe applies to entimines seen in Colombian collections and might not reflect the entire variation across tribes and genera. In most cases, there is more than one morphospecies with-
in each genus at each collection. Species were not identified. There remain specimens recognised as entimines, but not identified to tribe. Relevant information for entimine tribes represented in Colombia was compiled and is presented here. A general pictorial key is provided here just as a way to narrow down identifications (Fig. 2). Preliminary identifications using this key should be checked against the lists of characters offered for each tribe for confirmation.

Figures

Line drawings were produced by tracing over photographs into drawing software. Figures are grouped to show morphological variation or to keep relevant tribal information together. Maps are based on data from collections. The record density map (Fig. 3) was created using R version 3.5.2 (R Core Team 2018), interfaced through RStudio version 1.1.463 (RStudio Team 2016) with the R package ‘tmap’ (Tennekes 2018). Distribution maps in Figs 3–10 were created using SimpleMappr (Shorthouse 2010); each map is accompanied by sketches of the general appearance of the head in dorsal view of some representative specimens of the tribe as seen in Colombian collections.

Results

There are eight tribes, 50 genera, and 224 species of Entiminae recorded from Colombia. Seven genera and 142 species are considered endemic to the country (Wibmer and O’Brien 1986a; Morrone 1999; Girón and Cardona-Duque 2018; see species list, Suppl. material 1 or Girón 2020 – https://doi.org/10.15472/jdwfao). A general pictorial key to recognise these eight tribes is presented in Fig. 2. The list of species with their distribution in Colombia is presented. The genera *Pandeleteius* Schönherr, 1834 (Schönherr 1834, 129; Tanymecini, 44 spp.), *Compsus* Schönherr, 1823 (Schönherr 1823, col. 1140; Eustylini, 39 spp.), *Hypsonotus* Germar, 1824 (Germar 1824, 367; Lordopini, 17 spp.) and *Exorides* Pascoe, 1881: 43 (Pascoe 1881, 43; Eustylini, 14 spp.) are the most diverse in the country (numbers in parentheses indicate numbers of species recorded from Colombia).

Diversity by department

A total of 749 records of entimines were obtained from the revised collections (Suppl. material 2), with 248 (33%) from Valle del Cauca, followed by Magdalena (75, 10%), Antioquia (62, 8.3%), Risaralda (48, 6.4%), Boyacá (47, 6.3%), Nariño (35, 4.7%), Bolívar (31, 4.1%), Meta (27, 3.6%), Quindío (22, 3%), Cauca, Cundinamarca and Amazonas, each with 18 records (2.4%), followed by Tolima (11, 1.5%), Chocó (9, 1.2%), Santander (7, 1%), Putumayo and Vaupés with 6 each (0.8%), Norte de Santander (5, 0.7%), Vichada (3, 0.4%), Guaviare (2, 0.3) and Cesar with only one record (0.1%). The Departments of Atlántico, Caquetá, Córdoba, Huila, La Guajira, Sucre, Arauca, Guainia and San Andres, Providencia and Santa Catalina have no entimine occurrences from the studied collections (Fig. 2).
Figure 2. Pictorial key for the recognition of Tribes represented in Colombia. Preliminary identifications achieved using this key should be checked against the lists of characters offered for each tribe for confirmation.

Tribe Anypotactini Champion, 1911

Fig. 4

Recognition. Usually small (approx. 4–11 mm); scale coverage composed by appressed, mostly non-overlapping scales, usually uniform (in density and colour), usually brown (some species iridescent green or blue); usually with thick and erect
Figure 3. Record density map of Entiminae in Colombia. The colour gradient in the map represents the number of records by Department recorded from entomological collections (see Materials and Methods). Departments in white indicate no records and Departments increasingly darker with larger numbers of records (numbers in parentheses): Valle del Cauca (248), Magdalena (75), Antioquia (62), Risaralda (48), Boyacá (47), Nariño (35), Bolívar (31), Meta (27), Quindío (22), Cauca, Cundinamarca and Amazonas, (18 each), Tolima (11), Chocó (9), Santander (7), Putumayo and Vaupés (6 each), Norte de Santander (5), Vichada (3), Guaviare (2) and Cesar (1). Atlántico, Caquetá, Córdoba, Huila, La Guajira, Sucre, Arauca, Guainía and San Andrés, Providencia and Santa Catalina have no entimine occurrences from the studied collections. The data used to create this map are available in Suppl. material 2.
scale-like setae, rather uniformly distributed all over the surface; head (including rostrum) subrectangular [e.g. some *Anypotactus* Schönherr, 1840 (Schönherr 1840b, 299; Fig. 4B) and *Polydacrys* Schönherr, 1834 (Schönherr 1834, 130; Fig. 4D)] to conical [*Phanasora* Pascoe, 1881 (Pascoe 1881, 38; Fig. 4C), *Prepodellus* Kirsch, 1868 (Kirsch 1868, 239) and some *Anypotactus*], usually longer than wide; surface of rostrum flat, depressed or undulate, often with longitudinal carinae (Fig. 4B–D); nasal plate well developed (see Fig. 1B, C), forming an elevated, large and smooth plate in *Polydacrys* (Fig. 4D); mentum not covering the maxillae completely (Fig. 1G; except in *Polydacrys*, see Fig. 1A); antennal scape slender, at rest passing well below eye, often reaching or surpassing anterior margin of prothorax; anterior margin of prothorax in lateral view straight to oblique, never forming postocular lobe, lacking postocular setae [except in *Hyphantus* Germar, 1824 (Germar 1824, 334)]; elytral shoulders and hindwings weakly- to well-developed (Fig. 1N–P); legs relatively slender, femora clavate, often toothed (Fig. 1K).

Anypotactines can be confused with some small Naupactini, from which they can be recognised by the shape of the apical region of the metatibia, which is rectangular and fringed by flat spines along ventral and posterior margins in Naupactini (Fig. 1M, rounded in Anypotactini, see Fig. 1L). The general appearance of some Anypotactini resembles members of Tanymecini (Fig. 1D, with postocular setae) and can be differentiated, most of the time, by the lack of postocular setae in Anypotactini.

**Diversity.** The tribe contains 11 extant genera with 81 species described, all distributed in the New World (Wibmer and O’Brien 1986a, 44, Alonso-Zarazaga and Lyal 1999, 145). Seven genera represented by 11 species have been recorded from Colombia. *Hypsometopus* Kirsch, 1868 (Kirsch 1868, 223), represented by a single species, is considered endemic to the country; seven additional anypotactine species are considered endemic as well.

**General distribution.** The tribe ranges from south-western USA to Argentina and Chile and some Caribbean islands, including the following countries: Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, Peru, Paraguay, Uruguay, Venezuela; Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Cuba, Dominica, Granada, Guadeloupe, Puerto Rico, Saint Vincent; SW USA (Texas) (Alonso-Zarazaga and Lyal 1999). Most genera are distributed in Central America and northern South America, but the largest genus, *Hyphantus* Germar, 1824 (Germar 1824, 334; 45 species; see Vaurie 1963), is distributed in southern Brazil, Argentina, Paraguay and Uruguay.

**Distribution in Colombia.** Within the revised collections, anypotactines were recorded from Bolívar, Boyacá, Caldas, Cundinamarca, Magdalena, Quindío, Risaralda, Santander and Valle del Cauca (Fig. 4A). *Anypotactus* and *Polydacrys* were the most commonly found anypotactine genera in the revised collections; other anypotactines were found but remain identified to tribe only (Suppl. material 2; Fig. 4A).

**Bibliographic resources.** The best resource available for the tribe as a whole is the Biologia Centrali-Americana (Sharp and Champion 1911, 215, as ‘Anypotactina’). It includes a brief diagnosis of the tribe and diagnoses of the Central American
Figure 4. Distribution and some head variants of Anypotactini: A distribution map of records for Anypotactus, Bothynodontes, Phanasora, Polydacrys, Prepodellus and undetermined Anypotactini in Colombia B–D head in dorsal view: B Anypotactus C Phanasora D Polydacrys.

genera and species (with some keys). There is a key in van Emden (1944b, 506) to identify the tribe, as well as a key to genera (van Emden 1944b, 510). The only genus of the tribe that has been revised is Hyphantus (Vaurie 1963), but it is not represented in Colombia.

Natural history. Except for a few plant-association records for Hyphantus (Vaurie 1963) and records of an unidentified species of Bothynodontes on Alnus jorulensis (Betulaceae) in Caldas and Baccharis sp. (Asteraceae) in Tolima (Bustillo and Villegas Isaza 1986), the natural history of Anypotactini remains largely unknown.

Remarks. A large proportion of anypotactine records in Colombia are concentrated in the mountainous regions (Fig. 4A). Some specimens amongst the undetermined
Anypotactini from Caldas (Manizales, Río Blanco; MUSENUV) likely belong to an undescribed genus (Dr. Robert Anderson, pers. comm.). Specimens recorded from lowlands, especially from the north coast, share affinities with material seen from Central America at ASUCOB. One species of *Anypotactus* was included in a morphology-based phylogenetic analysis (Girón and Franz 2010), resulting as sister to *Apodrosus* Marshall, 1922 (Marshall 1922a, 59; Polydrusini, exclusively Caribbean); no anypotactines have been included in molecular-based phylogenetic analyses to date.

**Tribe Entimini Schönherr, 1823**

*Fig. 5*

**Recognition.** Medium to large (approx. 7–45 mm), often very robust; scale coverage either nearly uniform and dense or forming patches across surface, highly variable in colouration including brown, white, purple and iridescent tones of green and blue; head (including rostrum) subrectangular, usually longer than wide (Fig. 5B, C); rostrum usually apically broadened (Fig. 5C); anterior margin of prothorax in lateral view strongly sinuate forming conspicuous postocular lobe (Fig. 5D), often with fringe of postocular setae (Fig. 5D); elytral shoulders well developed, sometimes projected from general outline of body; hindwings well-developed; surface of elytra may be granulate or bear protuberances or tubercles.

Members of Entimini are generally easily distinguishable by their usually large size and the general bulky shape of their bodies [*Cydianerus* Schönherr, 1840 (Schönherr 1840a, 737), *Entimus, Polyteles* Germar, 1829 (Germar 1829, 358)] or the characteristic shape of their heads with narrow frons and rostrum with lateral margins apically diverging [*Cydianerus, Rhigus* Schönherr, 1823 (Schönherr 1823, col. 1138)]. A more detailed diagnosis is offered by Vaurie (1951).

**Diversity.** The tribe contains seven Neotropical genera with 46 species described to date, two of them fossil (Wibmer and O’Brien 1986a, 99, Gaiger 2001; Morrone 2002; Vanin and Gaiger 2005; Poinar et al. 2017). Three genera, represented by four species, have been recorded from Colombia, one of them fossil (Poinar et al. 2017).

**General distribution.** The tribe ranges from Mexico to Argentina, with no representatives in the Caribbean islands. Entimini can be found in the following countries: Argentina, Bolivia, Brazil, Colombia, Ecuador, French Guiana, Guyana, Paraguay, Peru, Uruguay, Venezuela; Costa Rica, El Salvador, Honduras, Mexico, Nicaragua, and Panama (Alonso-Zarazaga and Lyal 1999). All genera are mainly represented in Brazil.

**Distribution in Colombia.** In Colombian collections, representatives of Entimini (genus *Rhigus*) have been recorded from Amazonas and Tolima (Girón and Cardona-Duque 2018, 181). The distribution in the country for the rest of Colombian Entimini has not been recorded in literature. There is one record of the genus *Cydianerus* from Sucre in iNaturalist (https://www.inaturalist.org/observations/39591579).
Bibliographic resources. Entimini is one of the better-studied groups of Neotropical broad-nosed weevils. Sharp and Champion (1911, 300), with emphasis on Central American fauna, offer some diagnostic features for a wider concept of Entimini (as ‘Entimina’) which includes some genera currently placed elsewhere. Bovie (1908a, 2) offered a key to genera including illustrations (Bovie 1908a, plate 9). Morrone (1999) treated ‘Entimina’ in a broader sense, including ‘Entimina,’ ‘Lordopina,’ ‘Hypsonotina,’ ‘Promecopina’ and ‘Eudiagogina’ of Wibmer and O’Brien (1986a). Some genera of the tribe have been revised: *Entimus* (Vaurie 1951; Morrone 2002), *Rhigus* (Gaiger 2001) and *Phaedropus* Schönherr, 1823 (Schönherr 1823, col. 1140; Vanin 1983; not recorded from Colombia). A morphology-based phylogeny, key to the current genera and details on their morphology were presented by Vanin and Gaiger (2005).
Remarks. Even though members of Entimini are amongst the largest and most striking weevils in the subfamily, they are extremely rare in Colombian collections; only a few specimens of *Rhigus* have been found in the revised collections along with a few other unidentified Entimini. Romo and Morrone (2011) analysed biogeographic trends in the tribe. With the exception of the life cycle for *Entimus nobilis* (Olivier, 1790) (Olivier 1790, 525, Bruch 1932), the natural history of members of Entimini remains largely unknown. Plant associations have only been recorded for one species of *Phaedropus* (Vanin 1983) and one species of *Rhigus* (Gaiger 2001). There is one fossil species of *Cydianerus* recently described from Colombia (Poinar et al. 2017) and one fossil species of *Entimus* described from the USA (Scudder 1876). There is a recent study on the morphological variation and sexual dimorphism in *Cydianerus latruncularius* (Perty, 1832) (Perty 1832, 70, Regueira et al. 2020). *Rhigus speciosus* has been recently recorded from Colombia (Girón and Cardona-Duque 2018) and Peru (Gillett and Barr 2020). The type species for *Entimus, Entimus imperialis* Forster, 1771 (Forster 1771, 34), has been the focus of studies regarding the structural colouration of its scales which, as in other entimines, contain three-dimensional photonic crystals with diamond-type structure (e.g. Wilts et al. 2012). *Polyteles* Germar, 1829 (Germar 1829, 358) has not been recorded from Colombia, but its presence in the south of the country is likely, given that the genus is distributed in Argentina, Bolivia, Brazil, Ecuador, Paraguay, Peru and Uruguay.

Tribe Eudiagogini LeConte, 1874

Fig. 6

Recognition. Small weevils (approx. 3–8 mm); scale coverage usually light to dark brown, forming patterns on dorsal surface; iridescent areas with blue or green scales or reddish or yellowish stripes may be present in some genera; setae along surface may also be present; surface of body smooth and even; head (including rostrum) subrectangular, usually nearly as long as wide (Fig. 6B, C); rostrum robust, usually nearly parallel-sided (Fig. 6B, C); nasal plate (see Fig. 1B, C) usually well developed; anterior margin of prothorax in lateral view strongly sinuate forming conspicuous broad postocular lobe (Fig. 6D); elytral shoulders moderately to well-developed (e.g. Fig. 1O, P); legs relatively stout, femora never toothed [except in *Eurysaces* Schönherr, 1840 (Schönherr 1840b, 313), not present in Colombia].

Eudiagogines are easily recognisable by their small size, well-developed postocular lobes and overall robustness. They could potentially be confused with any-potactines because of their size, but their postocular lobe sets eudiagogines apart. They can resemble some small Entimini, but eudiagogines have a comparatively shorter and stouter rostrum. Additional characters to define the tribe can be found in Lacordaire (1863, 384, in French).

Diversity. The tribe contains ten extant genera with 100 species (including two fossil species in Dominican amber; Poinar and Brown 2011; Poinar and Legalov 2017)
Broad-nosed weevils of Colombia (Coleoptera, Curculionidae, Entiminae)

Figure 6. Distribution and some head variants of Eudiagogini: A distribution map of records for *Promecops* and undetermined eudiagogines in Colombia B, C head in dorsal view: B *Promecops* C *Colecerus* D head and anterior section of prothorax in lateral view of *Promecops* showing well-developed postocular lobe.

described to date, all distributed in the New World (as ‘Promecopini’ in Wibmer and O’Brien 1986a, 81, O’Brien and Kovarik 2001). Two genera, *Colecerus* Schönherr, 1840 (Schönherr 1840a, 927) and *Promecops* Sahlberg, 1823 (Sahlberg 1823, 30), represented by six species, have been recorded from Colombia; four species are endemic.

General distribution. The tribe ranges from the USA to Argentina, including some Caribbean islands. Eudiagogini can be found in the following countries: Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, French Guiana, Guyana, Paraguay, Peru, Uruguay, Venezuela; Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Grenadines, Guadeloupe, St. Vincent; NC, NE, SE, SW USA (Alonso-Zarazaga and Lyal 1999).
Distribution in Colombia. Representatives of the tribe have been recorded in collections from Amazonas, Bolívar, Chocó, Magdalena, Meta, Risaralda, Tolima, Valle del Cauca, and Vaupés (Suppl. material 2, Fig. 6A).

Bibliographic resources. The original diagnosis for Eudiagogini (then named Promécopides) was given by Lacordaire (1863, 384, in French). In the Biologia Centrali-Americana (Sharp and Champion 1911, 300) eudiagoge genera were considered part of an extended concept of Entimini (as ‘Entimina’). Morrone (1999) treated ‘Eudiagogina’ and ‘Promecopina’ as part of ‘Entimina’. A revision of Eudiagogus Schönherr, 1840 (Schönherr 1840b, 307), was published by Warner (1979). LeConte and Horn (1876, 108) treated the genera recorded from North America. Voss (1934, 74, in German, as ‘Promecopina’), provided a key to genera (containing genera of Entimini) and keys to species for some eudiagoge genera.

Remarks. Promecops contains over 60% of the species and is also the most widespread genus of the tribe; some species are considered economically important in soybean cultivars in Argentina (Lázaro et al. 1997, 1998) and Brazil (Rocha Barreto and Cavalet 2016). One species of Colecerus has been associated with cultivated Pecan trees in Mexico (Soto-Hernández and Barros-Barrios 2018). In North America, Eudiagogus has been associated with legumes in the genus Sesbania (Warner 1979); Eudiagogus has not been recorded from Colombia, but its presence in the south of the country is likely, given that the genus is distributed in Argentina, Bolivia, Brazil, Ecuador, Paraguay, Peru, Uruguay, Costa Rica, Honduras, Mexico and USA. Promecops is the only eudiagoge genus recorded from the revised Colombian collections; other eudiagonines were located, but remain undetermined (Suppl. material 2, Fig. 6A).

Tribe Eustylini Lacordaire, 1863
Fig. 7

Recognition. Medium to large weevils (approx. 10–25 mm); scale coverage highly variable in presence, density and colouration; iridescent scales, erect setae or waxy secretions are frequently present; surface smooth and even or strongly sculptured and irregular; head (including rostrum; Fig. 7C–F) subrectangular, nearly as long or longer than wide; eyes small to mid-sized, slightly dorsally positioned; frons usually as wide as or narrower than interantennal distance, often bearing median fovea (see Fig. 1C); rostrum nearly parallel-sided or broadened apically; dorsal surface of rostrum with variable elevations or depressions, including longitudinal carinae or oblique fossae; antennal scrobe generally fully visible in dorsal view (Fig. 7C–F); nasal plate (see Fig. 1B, C) usually well developed, either depressed, flat or elevated regarding surface of rostrum; anterior margin of prothorax in lateral view straight, seldom slightly sinuate (Fig. 1E), never forming conspicuous postocular lobe; postocular setae may be present, if so, forming a fringe instead of a tuft (compare Fig. 1D vs. 1F); elytral shoulders usually well-developed, absent in Brachyomus La-
cordaire, 1863 (Lacordaire 1863, 130), reduced in some *Compsus* and *Exorides* Pascoe, 1881 (Pascoe 1881, 43) (see Fig. 1N–P); tubercles and apical projections may be present on elytra; femora usually not toothed (Fig. 1H; except in some *Eustylus* Schönherr, 1842 (Schönherr 1842, 40), Fig. 1I).

Eustylines are relatively easy to recognise amongst South American entimines by their size, frequently iridescent/bright colourations and the shape of their head (Fig. 7C–F). They may be confused with similarly-coloured Naupactini, but the shape of the head, eyes, nasal plate and apex of metatibia [compare Fig. 1L (Eustylini) vs. 1M (Naupactini)] differentiate them.

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**Figure 7.** Distribution and some head variants of Eustylini: **A, B** distribution maps of Eustylini in Colombia: **A** records for *Brachyomus*, *Compsus*, *Eustylus* and *Exophthalmus* **B** records for *Exorides*, *Oxyderces*, *Synthlibonotus*, *Xestogaster* and undetermined eustylines **C–F** head in dorsal view: **C** *Exophthalmus* **D** *Synthlibonotus* **E** *Exorides* **F** *Eustylus* with characteristic broad antennal scape.
Diversity. Eustylini contains 25 genera with 336 species described to date. Eight genera, represented by 73 species, have been recorded from Colombia, 52 of which are endemic.

General distribution. Eustylini ranges from south-western USA to Argentina, with its highest diversity in the Caribbean, Central America and northern South America, including the following countries: Argentina, Bolivia, Brazil, Colombia, Ecuador (incl. Galapagos Islands), French Guiana, Guyana, Paraguay, Peru, Surinam, Trinidad, Venezuela; Belize, Costa Rica (incl. Cocos Island), El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Antigua, Barbados, Cuba, Dominica, Dominican Republic, Guadeloupe, Haiti, Hispaniola, Jamaica, Martinique, Montserrat, Nevis, Puerto Rico (incl. Mona Is., Vieques Is.), St. Barthelemy, St. Croix, St. Kitts, St. Lucia, St. Thomas, St. Vincent; NC, NE, SE, SW USA (Alonso-Zarazaga and Lyal 1999).

Distribution in Colombia. Representatives of the tribe have been recorded from 21 Departments in the Country: Amazonas, Antioquia, Bolívar, Boyacá, Caldas, Casanare, Cauca, Chocó, Cundinamarca, Magdalena, Meta, Nariño, Norte de Santander, Putumayo, Quindío, Risaralda, Santander, Tolima, Valle del Cauca, Vau- pés and Vichada (Suppl. material 2).

Bibliographic resources. The Biologia Centrali-Americana (Sharp and Champion 1911) may be a good place to start understanding the diversity of Eustylini (as ‘Exophthalmina’, p. 247 and ‘Playomina’, p. 282). There is a key to identify species of Brachyomus in Faust (1893, 16, in German). Marshall (1922b) discusses several classification issues with Compsus, Exophthalmus Schönherr, 1823 (Schönherr 1823, col. 1140) and allies, describes new genera and species and presents a key to identify species of Exorides (p. 203). Hustache (1938a, in French) revised part of South American Compsus; O’Brien and Peña (2012) translated to English and modified Hustache’s key and provided identification and records for two species of Compsus recorded as Citrus pests in Colombia. Franz (2010a) re-described several type species in Eustylini and presented a morphology-based phylogeny of the so-called “Exophthalmus genus complex”, which involves several eustyline genera, especially Central American and Caribbean groups. Girón and Chamorro (2020) defined the “Compsus genus complex” and discussed affinities amongst species in this group. Some eustylines were included in phylogenetic analyses emphasising South American taxa by Marvaldi et al. (2018).

Remarks. The circumscription of Eustylini has been problematic since Lacordaire (1863), in French (some as ‘Cyphides’ p. 107, some as ‘Geonémides’ p. 125 and some as ‘Eustylides’ p. 205). Different genera, currently placed in Eustylini, have also been grouped together with members of Phyllobiini and Naupactini through time; most genera were treated as incertae sedis in the Coleopterorum Catalogus (Lona 1938: 508–525, 530–532). Kuschel (in Wibmer and O’Brien 1986a) brought together the majority of the genera that now compose Eustylini into a single tribe; the tribe is treated as ‘Eustylinina’ by Morrone (1999). Some additions to the tribe have been made with transfers from other tribes, justified by a morphology-based
phylogeny presented by Franz (2012). Marvaldi et al. (2018) made the most recent addition by placing Galapagonotus Anderson & Lanteri, 2000 (Anderson and Lanteri 2000, 3) in Eustylini; they indicate that Coconotus Anderson & Lanteri, 2000 (Anderson and Lanteri 2000, 6) might also be an eustyline, but this remains to be confirmed, as it also shares features with the geonemine genus Lachnopus Schöherr, 1840 (Schöherr 1840b, 380), particularly with species in the luctuosus species-group (see Girón et al. 2018). Recent phylogenetic analyses (Franz 2012; Zhang et al. 2017; Marvaldi et al. 2018) indicate close relationships of Eustylini with taxa currently placed in Geonemini Gistel, 1856 (Gistel 1856, 373), which is primarily Caribbean and Central American.

The generic limits between some eustyline genera are not clearly defined; this is particularly true for Compsus, Oxyderces Schöherr, 1823 (Schöherr 1823, col. 1140), Exorides and Xestogaster Marshall, 1922 (Marshall 1922b, 221), all well represented in Colombia. This highlights the pressing need for taxonomic and phylogenetic revisions in the tribe, including (ideally recent) Colombian material that is already deposited in national collections. This particular group is part of the “Compsus genus complex” (Girón and Chamorro 2020); the authors compare several species of Compsus, Oxyderces and Exophthalmus which, by their colouration, may be confused with the golden-headed weevil, Compsus auricephalus (Say, 1824) (Say 1824), which is the only species of Compsus to occur in North America.

Members of Eustylini are amongst the most commonly found entimines in Colombian biological collections; Compsus is the most frequently observed and collected genus, followed by Exorides and perhaps Eustylus. Species of Exophthalmus are rare and seem to be restricted to the Pacific Region. Species of Compsus have been recorded as Citrus pests (Cano et al. 2002a, b; Gallego et al. 2012, O’Brien and Peña 2012) in Colombia and sugar cane in Venezuela (Kuschel 1955a). The Colombian endemic Oxyderces viridipes (Boheman in Schöherr 1840b, 179) has been intercepted at ports in the USA on Hydrangea flowers (Hydrangeaceae; see Girón and Chamorro 2020). In nature, eustylines commonly rest with their prolegs extended towards the front of the body.

Tribe Lordopini Schöherr, 1823
Fig. 8

Recognition. Medium-sized weevils (approx. 10–20 mm); scale coverage variable in presence, density, pattern and colouration; iridescent scales and setae may be present; surface usually even (not coarsely sculptured); head (including rostrum, Fig. 8B, C) subrectangular to conical, usually longer than wide; eyes usually large and elongated in lateral view, weakly to moderately projecting from surface of head; frons nearly as wide as or wider than interantennal distance, often bearing median fovea; rostrum nearly parallel-sided, broadened apically (Fig. 8B) or with lateral margins apically converging (Fig. 8C); dorsal surface of rostrum either flat
or with one or three longitudinal carinae; antennal scrobe generally only visible at apical region in dorsal view; nasal plate (see Fig. 1B, C) usually well developed, flat or depressed regarding surface of rostrum; anterior margin of prothorax in lateral view strongly sinuate, forming conspicuous postocular lobe (Fig. 8D); postocular setae present, forming a fringe instead of a tuft (Fig. 8D); elytral shoulders absent to well-developed (see Fig. 1N–P); tubercles may be present on elytra; legs relatively slender.

Amongst entimines with well-developed postocular lobe, lordopines are recognisable because of their elongated rostrum (Fig. 8B, D) or their conical heads with apically-converging lateral margins of the rostrum (Fig. 8C). They may be...
confused with mid-sized Entimini, but the bulky shape of the body of most Entimini and the overall shape of the head differentiate them (compare Fig. 5B, C with Fig. 8B, C).

Diversity. There are 44 genera with 291 species of Lordopini described (Wibmer and O’Brien 1986a). Eleven genera, represented by 33 species, have been recorded from Colombia, four and 25, respectively, being endemic.

General distribution. Lordopini is a strictly Neotropical tribe, ranging from Mexico to Paraguay, including some of the Lesser Antilles. Lordopies have been recorded from the following countries: Argentina, Bolivia, Brazil, Colombia, Ecuador, French Guiana, Paraguay, Peru, Venezuela; Costa Rica, Belize, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Grenada, St. Vincent (Alonso-Zarazaga and Lyal 1999). The higher diversity of this tribe is massively concentrated in Brazil.

Distribution in Colombia. Representatives of Lordopini have been recorded from Bolívar, Magdalena, Nariño, Quindio, Santander, Valle del Cauca and Vaupés (Suppl. material 2).

Bibliographic resources. A general diagnosis for Lordopini can be found in Lacordaire (1863, 260, as ‘Hypsonotides’, in French). Only two genera appear in the Biologia Centrali-Americana (Sharp and Champion 1911, 300, under ‘Entimina’). Kessel (1932, 1935, 1937, in German) revised several genera and provided keys to genera and species, focusing on Brazilian fauna, but these volumes are not yet available online and are rather difficult to find in libraries.

Remarks. Lordopini is one of the least-studied entimine tribes in the region and needs systematic revision: 18 genera are Brazilian endemics; more than one third of the species belong to Hypsonotus Germar, 1824 (Germar 1824, 367, 115 spp.) and one fifth belongs to the genus Lordops Schönherr, 1823 (Schönherr 1823, col. 1142, 59 spp.). From the remaining genera, 17 are monotypic and 11 are represented by only two species. All of these facts might indicate problems with current generic definitions. Furthermore, there are no phylogenetic analyses, including lordopines, to date. There are records of an undetermined species of Hypsonotus attacking peanuts in Brazil (Araujo et al. 1977); other than that, the natural history of the Lordopini remains largely unknown.

Tribe Naupactini Gistel, 1856

Fig. 9

Recognition. Small to medium weevils (approx. 3.5–35 mm); scale coverage highly variable in presence, density and colouration; iridescent scales, erect setae or waxy secretions are frequently present; surface usually even; head (including rostrum; Fig. 9C–F) subrectangular to trapezoid, nearly as long or shorter than wide; eyes rounded to oval in lateral view, small to large, laterally positioned and moderately to strongly projected from surface of head; frons usually as wide as or wider than interantennal distance, often bearing median longitudinal furrow extending through...
rostrum; rostrum with lateral margins usually apically converging or nearly parallel-sided, sometimes slightly broadened apically (Fig. 9C–F); dorsal surface of rostrum usually flat (excepting median furrow; broadly anteriorly depressed in *Platyomus* Sahlberg, 1823 (Sahlberg 1823, 29) and relatives, for example, Fig. 9F); antennal scrobe generally not broadly visible in dorsal view; nasal plate (see Fig. 1B, C) of variable development, usually flat to depressed regarding surface of rostrum; anterior margin of prothorax in lateral view straight, never forming postocular lobe; postocular setae absent; elytral shoulders absent to well-developed (see Fig. 1N–P); tubercles may be present on elytra; profemora sometimes enlarged (see Fig. 1J); apical region of metatibia rectangular, fringed by spines along ventral and posterior margins (Fig. 1M).

Naupactines are relatively easy to recognise amongst South American entimines by the overall shape of their head, the usual presence of a median longitudinal furrow along the head and rostrum and the characteristic apex of their metatibia. They may be confused with similarly-coloured Eustylini, but the overall shape of the head (compare Fig. 7C–F with Fig. 9C–F) and the apex of the metatibia differentiate them (compare Fig. 7C–F with Fig. 9C–F). Small naupactines may be confused with anypotactines, from which they can be differentiated by the metatibial apex. From *Tanymecini*, naupactines can be distinguished by the lack of postocular setae in *Naupactini* and the shape of the metatibial apex.

**Diversity.** Naupactini contains 67 genera with over 500 species described (Lanteri and del Río 2016a). Fourteen genera, represented by 36 species have been recorded from Colombia, two genera and 22 species being endemic.

**General distribution.** In the New World, Naupactini ranges from the USA to Argentina and Chile, including some of the Caribbean islands. Naupactines have been recorded from the following countries: Argentina, Bolivia, Brazil, Chile (including Isla de Pascua, Juan Fernández Is.), Colombia, Ecuador (including Galapagos Is.), French Guiana, Guyana, Paraguay, Peru, Uruguay, Surinam, Trinidad, Venezuela; Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; USA (Alonso-Zarazaga and Lyal 1999). Its highest diversity is concentrated in Argentina, Bolivia, Brazil, Paraguay and Uruguay, which coincides with areas where they have been extensively studied.

**Distribution in Colombia.** Representatives of the tribe have been recorded from 18 Departments: Antioquia, Bolívar, Boyacá, Caldas, Casanare, Cauca, Cesar, Chocó, Cundinamarca, Guaviare, Magdalena, Meta, Nariño, Quindío, Risaralda, Valle del Cauca, Vaupés and Vichada (Suppl. material 2).

Figure 9. Distribution and some head variants of Naupactini: A, B distribution maps of Naupactini in Colombia: A records for Amphideritus, Lanterius, Mimographus and Naupactus B records for Litostylus, Platyomus and undetermined naupactines C–F head in dorsal view: C Mimographus D undetermined Naupactini E Litostylus F “Platyomus” (see discussion under Naupactini) with characteristic broad and flattened antennal scape.

2012, 2013, 2019, 2020; Lanteri et al. 2017). There have also been recent efforts to elucidate the phylogenetic relationships amongst naupactine genera (Lanteri and del Río 2016a; del Río et al. 2018) and with other entimines (Marvaldi et al. 2018). Keys to genera can be found in Sharp and Champion (1911, 221) and van Emden (van Emden 1944b); a key to Andean genera is available in del Río and Lanteri (2011a).

Remarks. Naupactini is the largest tribe of New World entimines and one of the best known, mainly because of the revisionary and phylogenetic efforts of Dr. Analía Lanteri, Dr. Adriana Marvaldi, Dr. Guadalupe del Río and collaborators, based in Argentina. Different naupactine clades are specific to certain biogeographic regions in South America (see Lanteri 1992; del Río and Lanteri 2011a), with flightlessness
and parthenogenesis being frequent in certain systems (Lanteri and Normark 1995; Guzmán et al. 2012; Lanteri et al. 2013; Lanteri and del Río 2016a). A detailed study of the female genitalia and oviposition habits in Naupactini, in a phylogenetic context, can be found in Lanteri and del Río (2008).

Most revisionary efforts to date have focused on fauna from Argentina, Brazil and surrounding countries. In northern South America, naupactines from Venezuela were studied by Bordón [1991, genus Macrostylus Boheman, 1840 (Schönherr 1840a, 921), 26 spp. described as new; 1997, genus Naupactus Dejean, 1821 (Dejean 1821, 94), 24 spp. described as new]; none of these species has been recorded from Colombia to date. Other than that, the naupactine fauna from northern South America, Central America and the Caribbean is still in need of revision.

Several naupactine species are economically important (e.g. Lanteri et al. 2002, 2013); Leschenius vulcanorum (Kirsch, 1889) (Kirsch 1889, 17) has been recorded in Colombia as the ‘Potato shooter’ or ‘tiroteador de la papa’ (Canchala 1992; Peña 2001; Cortázar Gómez et al. 2012).

Members of Naupactini are very common in Colombian entomological collections, especially the genera Lanterius Alonso-Zarazaga & Lyal, 1999 (Alonso-Zarazaga and Lyal 1999, 164), Mimographus Schönherr, 1847 (Schönherr 1847, 34) and allies. Platyomus was recorded for the first time for Colombia by Girón (2007a; for Risaralda and Valle del Cauca) and then by Cardona-Duque et al. (2018; for Antioquia); these records might be erroneously identified and, instead, the specimens cited could be representatives of Chamaelops Kirsch, 1868 (Kirsch 1868, 235), which is fairly closely related to Platyomus (Lanteri and del Río 2016a); Chamaelops has previously been recorded for Colombia, considered endemic and includes two described species. Galapaganus Lanteri, 1992 (Lanteri 1992, 230) and Pantomorus Schönherr, 1840 (Schönherr 1840a, 942) were also recorded by Girón (2007a; both from Valle del Cauca) and are presented here (see Suppl. material 2; including records from Bolívar for Galapaganus and from Casanare and Meta for Pantomorus), but these records need to be confirmed.

Lanterius was erected by Alonso–Zarazaga and Lyal (1999, 164) to group species that were previously placed in a misidentified concept of Mimographus Schönherr, 1847 (Schönherr 1847, 34; see del Río and Lanteri 2018, 331). Even though del Río and Lanteri (2018) indicate that each of the Mimographus species, listed by Wibmer and O’Brien (1986a, 56), needs to be studied in detail in order to determine their appropriate placement, the Colombian species are here listed as new combinations under Lanterius in an attempt to prevent confusion, as there are species of true Mimographus recorded from Colombia as well. These new combinations are: Lanterius ardosiacus (Kirsch, 1868: 229), comb. nov., Lanterius cinereoguttatus (Champion, 1911: 228), comb. nov., Lanterius hirtus (Voss, 1953: 60), comb. nov., Lanterius laesicollis (Schönherr, 1847: 35), comb. nov., Lanterius micans (Kirsch, 1868: 229), comb. nov., Lanterius rotundicollis (Kirsch, 1889: 15), comb. nov., Lanterius versicolor (Kirsch, 1889: 15), comb. nov., and Lanterius vittatus (Kirsch, 1889: 14), comb. nov.
Tribe Premnotrypini Kuschel, 1956
Fig. 10D

**Recognition.** Small weevils (approx. 4–15 mm); scale coverage usually dense, uniform, and brown, often with patches of erect scales or setae; surface from even to undulate or tuberculate; head (including rostrum, Fig. 10D) usually conical and longer than wide; eyes laterally positioned and weakly to moderately projected from surface of head; frons wider than interantennal distance; rostrum elongate, usually with lateral margins apically converging, sometimes slightly broadened apically; dorsal surface of rostrum flat to undulate; mandibular scar reduced to absent; antennal scrobe generally not broadly visible in dorsal view; nasal plate (see Fig. 1B, C) of variable development, usually flat to elevated regarding surface of rostrum; anterior margin of prothorax in lateral view sinuate, forming well-developed and large postocular lobe (see Fig. 1F); postocular setae forming fringe; elytral shoulders usually well-developed (see Fig. 1P); tubercles may be present on elytra.

Premnotrypines may not be easily recognisable as entimines, as their rostrum is relatively slender and usually directed towards the body when mounted; the reduction or absence of mandibular scars would make them pass as other miscellaneous Curculionidae. Amongst entimines, they can be recognised by their small size and well-developed postocular lobe, fringed by postocular setae. By their size, premnotrypines can be confused with anypotactines or small tanymecines, but can be recognised by the presence of a well-developed postocular lobe, lacking in both Anypotactini and Tanymecini.

**Diversity.** There are only three genera in Premnotrypini with 28 species (Kuschel 1956; Wibmer and O'Brien 1986a, 88). There is only one species recorded from Colombia.

**General distribution.** Premnotrypini is an exclusively Andean group, distributed from Colombia to Bolivia and northern Chile. Their diversity is concentrated in Bolivia and Peru, at elevations over 3000 m a.s.l. No specimens of this tribe were identified in the collections revised for this work.

**Bibliographic resources.** The tribal diagnosis, key to genera, descriptions of two of the genera and keys to all species can be found in Kuschel (1956, in Spanish).

**Remarks.** The genus *Premnotrypes* is associated with potato cultivars and contains economically-important species commonly known as the “Andean potato weevils” (see Alcázar and Cisneros 1998). There are many studies in the natural history and control of pest species (e.g. Alcalá and Alcázar 1976; Calvache 1987; Dueñas 1989), but no phylogenetic studies in Premnotrypini have been published to date.

Tribe Tanymecini Lacordaire, 1863
Fig. 10A–C

**Recognition.** Small to large weevils (approx. 4–20 mm); scale coverage variable in presence, density and colouration, often dense and uniform, with colouration
patterns formed by scales on dorsal surface; surface usually even, sometimes granulate; head (including rostrum; Fig. 10B, C) usually subrectangular and nearly as long as wide; eyes laterally positioned and weakly to strongly projected from surface of head; frons usually as wide as interantennal distance, bearing median fovea or small (narrow and short) longitudinal furrow that may extend along rostrum; rostrum subquadrate, usually with lateral margins parallel, sometimes slightly constricted basad of antennal insertion; dorsal surface of rostrum flat to slightly depressed; antennal scrobe only visible along anterior section in dorsal view; epistoma usually well-developed; nasal plate (see Fig. 1B, C) of variable development, flat to depressed regarding surface of rostrum; anterior margin of prothorax in lateral view straight (Fig. 1D), never forming well-developed postocular lobe; postocular setae usually present, clearly visible, grouped as a tuft, usually positioned near ventral margin of prothorax (Fig. 1D); elytral shoulders absent to well-developed (see Fig. 1N–P); tubercles may be present on elytra and elytral apices may be projected; profemora frequently enlarged (see Fig. 1J).

Amongst Neotropical entimines, members of Tanymecini are easily recognizable by the presence of postocular setae, although not all tanymecines have this feature and not all the species with postocular setae are tanymecines. The postocular setae in Tanymecini are usually clustered, forming a tuft as opposed to distributed along the margin forming a fringe (compare Fig. 1D with Fig. 1F). Some tanymecines can be potentially confused with Naupactini because of their flat frons, median furrow and laterally-positioned eyes, but the presence of postocular setae in Tanymecini sets them apart easily; they can also be differentiated by the particular shape of the apex of the metatibia in Naupactini, which is rectangular and fringed by spines along the ventral and posterior margins (Fig. 1M; compare with rounded margins in tanymecines, Fig. 1L).

**Diversity.** There are about 100 genera of Tanymecini distributed worldwide. Eighteen genera are represented throughout the Americas by 325 species (O’Brien and Wibmer 1982, 45; Wibmer and O’Brien 1986a, 67; Alonso-Zarazaga and Lyal 1999, 178; Morrone 1999, 143; Cortés-Hernández and Anderson 2019; Cortés-Hernández and Morrone 2020). There are 58 species of Tanymecini recorded from Colombia, 29 of them being considered endemic.

**General distribution.** Tanymecines are distributed throughout the Americas and the Caribbean islands, including the following countries: Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, Peru, Paraguay, Uruguay, Venezuela; Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Bahamas, Cuba, Dominica, Dominican Republic, Grand Cayman, Grenada, Guadeloupe, Haiti, Jamaica, Saint Vincent; C, E Canada, USA (Alonso-Zarazaga and Lyal 1999). Their highest diversity is concentrated in Central America and northern South America.

**Distribution in Colombia.** Tanymecines have been found in collections from Antioquia, Bolívar, Boyacá, Caldas, Cauca, Cundinamarca, Magdalena, Meta, Narino, Norte de Santander, Quindío, Risaralda and Valle del Cauca (Suppl. material 2).
Remarks. Tanymecini is one of the best-known tribes of entimines of the Americas, thanks to the works of Anne Howden. Tanymecines, especially *Pandeleteius*, are quite common in Colombia. In nature, tanymecines are commonly seen with their prolegs resting perpendicular to the body axis, a type of behaviour shared with some naupactines.

*Figure 11.* Distribution of undetermined entimines in Colombia.
List of valid species of Entiminae recorded from Colombia

Genera and species in bold letters indicate endemic taxa. Fossil taxa are indicated by (+). Localities in bold letters indicate type locality. Localities marked with an asterisk are records taken from ASUCOB. A double asterisk means that identifications need to be corroborated. A question mark (?) in the distribution indicates that the locality record is doubtful and needs confirmation. Type localities from countries other than Colombia are omitted. Taxon names include author, year and page number of the original description (e.g. *Anypotactus* Schönherr 1840b, 299); for some of Schönherr’s taxa, the page number corresponds to a column number in the publication and is indicated by ‘col.’ (e.g. Entiminae Schönherr 1823, col. 1138). See other locality remarks in the methods section.

**Tribe Anypotactini Champion, 1911: 215**

Genus *Anypotactus* Schönherr, 1840: 299

Distribution: Bolivia, Colombia, Peru, Venezuela; Costa Rica, El Salvador, Guatemala, Honduras, Nicaragua, Panama.


*Anypotactus exilis* Boheman, 1840: 300

Distribution: Colombia, Venezuela; Costa Rica, Guatemala, Honduras, Nicaragua, Panama.


*Anypotactus morosus* (Boheman, 1840: 449)

Distribution: Colombia.

References: As *Polydrosus morosus* by Boheman in Schönherr 1840b, 449; Kuschel 1955b, 277 (as *Anisactus morosus*), Kuschel in Wibmer and O’Brien 1986a, 44; Morrone 1999, 135.

Genus *Bothinodontes* Kirsch, 1868: 241

Distribution: Colombia (Caldas, Tolima); Mexico (?).
Bothinodontes gibbipennis (Champion, 1911: 332)

Distribution: Colombia; Mexico (?).


Bothinodontes squalidus Kirsch, 1868: 243

Distribution: Colombia (‘Bogotá’).

References: Kirsch 1868, 243; Wibmer and O’Brien 1986a, 45; Morrone 1999, 135.

Genus Cylloproctus Faust, 1892: 22

Distribution: Brazil, Colombia, Venezuela.


Cylloproctus modestus Faust, 1892: 23

Distribution: Colombia [Columbia].

References: Faust 1892, 23 (in footnote), Wibmer and O’Brien 1986a, 45; Morrone 1999, 135.

Cylloproctus pantomoroides Voss, 1936: 109

Distribution: Colombia (Cundinamarca: Fusagasugá).


Genus Hypsometopus Kirsch, 1868: 222

Distribution: Colombia.

**Hypsometopus inquinatus** Kirsch, 1868: 223

Distribution: Colombia (‘Bogotá’).

Genus *Phanasora* Pascoe, 1881: 38

Distribution: Colombia, Ecuador.
References: Pascoe 1881, 38; Wibmer and O’Brien 1986a, 45; Alonso-Zarazaga and Lyal 1999, 146; Morrone 1999, 137.

**Phanasora plumbea** Pascoe, 1881: 39

Distribution: Colombia (‘Bogotá’).
References: Pascoe 1881, 39; Wibmer and O’Brien 1986a, 45; Morrone 1999, 137.

Genus *Polydacrys* Schönherr, 1834: 130

Distribution: Bolivia, Colombia, Cuba, Dominica, Granada, Guadeloupe, Puerto Rico, Saint Vincent; Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; SW USA (Texas).

**Polydacrys seriegranosus** Champion, 1911: 218

Distribution: Bolivia, Colombia; Panama.

Genus *Prepodellus* Kirsch, 1868: 239

Distribution: Colombia; Costa Rica, Mexico, Nicaragua, Panama.
Prepodellus nigriclavis Kirsch, 1868: 240

Distribution: Colombia (‘Bogotá’, Cundinamarca (La Vega*)).
References: Kirsch 1868, 240; Wibmer and O’Brien 1986a, 44; Morrone 1999, 137.

Prepodellus ruficornis Kirsch, 1868: 240

Distribution: Colombia (‘Bogotá’, Cundinamarca (Mosquera*)).
References: Kirsch 1868, 240; Wibmer and O’Brien 1986a, 44; Morrone 1999, 137.

Tribe Entimini Schönherr, 1823: col. 1138

Genus Cydianerus Schönherr, 1840: 737

Distribution: Argentina, Bolivia, Brazil, Colombia, Paraguay, Venezuela; Costa Rica, El Salvador, Honduras, Mexico, Panama.
References: Schönherr 1840a, 737; Sharp and Champion 1911, 300 as Entimina, O’Brien and Wibmer 1982, 67; Wibmer and O’Brien 1986a, 101; Alonso-Zarazaga and Lyal 1999, 144; Morrone 1999, 110; Poinar et al. 2017 (key to species).

Cydianerus eukrinus Poinar, Bukejs & Legalov, 2017: 88 (+)

Distribution: Colombia (Santander, copal resin).

Cydianerus pascoei (Bovie, 1908a: 4)

Distribution: Colombia; Panama.

Genus Entimus Germar, 1817: 341

Distribution: Argentina, Bolivia, Brazil, Colombia, Ecuador, French, Guiana, Guyana, Paraguay, Peru, Uruguay; Costa Rica, Nicaragua, Panama.
Entimus arrogans Pascoe 1872: 448

Distribution: Colombia; Costa Rica, Nicaragua, Panama.

Entimus granulatus (Linnaeus, 1758: 386)

Distribution: Bolivia, Brazil, Colombia, Ecuador, French Guiana, Guyana, Peru; Panama.

Genus Rhigus Schönherr, 1823: col. 1138

Distribution: Argentina, Brazil, Colombia, French Guiana, Paraguay, Peru.

Rhigus speciosus (Linnaeus, 1758: 385)

Distribution: Brazil, Colombia (Amazonas, Tolima), French Guiana, Peru.
References: Linnaeus 1758, 385 (as Curculio speciosus), Wibmer and O’Brien 1986a, 102; Morrone 1999, 121; Gaiger 2001, 63; Girón and Cardona-Duque 2018, 181 (first record for Colombia), Gillett and Barr 2020 (first record for Peru).

Tribe Eudiagogini LeConte, 1874: 454

Genus Colecerus Schönherr, 1840: 927

Distribution: Colombia; Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua; SW USA.

Colecerus albidus (Chevrolat, 1881: XXXVIII)

Distribution: Colombia (‘Bogotá’).

Genus *Promecops* Sahlberg, 1823: 30

Distribution: Argentina, Bolivia, Brazil, Colombia, Ecuador, French Guiana, Guyana, Paraguay, Peru, Venezuela; Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Grenadines, Guadeloupe, St. Vincent.


*Promecops lepidoides* Voss, 1934: 75

Distribution: Colombia (Meta: **Villavicencio**).

References: Voss 1934, 75 (in key), Wibmer and O’Brien 1986a, 83; Morrone 1999, 120.

*Promecops lepidus* Fåhraeus, 1840: 317

Distribution: **Colombia**, [Orinoco] (Magdalena (Ciénaga)*).


*Promecops leucothyreus* Fåhraeus, 1840: 328

Distribution: Bolivia, Colombia (‘**Bogotá**’, Antioquia (Puerto Berrio), Bolívar (Cartagena)*, Magdalena); Guatemala, Honduras, Mexico, Nicaragua.

References: Fåhraeus in Schönherr 1840b, 328; Voss 1934, 80 (in key, as *Promecops leucothyrea*), O’Brien and Wibmer 1982, 59; Wibmer and O’Brien 1986a, 83; Morrone 1999, 120.

*Promecops rhombicus* Fåhraeus, 1840: 326

Distribution: **Colombia** [Columbia].

References: Fåhraeus in Schönherr 1840b, 326; Voss 1934, 97 (listed as *Promecops rhombica*), Wibmer and O’Brien 1986a, 83; Morrone 1999, 120.
Promecops rhombifer Fåhraeus, 1840: 325

Distribution: **Colombia**, Guyana, Venezuela.

References: Fåhraeus in Schönher 1840b, 325; Voss 1934, 90 (in key, as Promecops rhombifera), Wibmer and O’Brien 1986a, 83; Morrone 1999, 120.

**Tribe Eustylini Lacordaire, 1863: 205**

Genus *Brachyomus* Lacordaire, 1863: 130

Distribution: Colombia, Ecuador, French Guiana, Peru, Trinidad, Venezuela; St. Vincent.

References: Lacordaire 1863, 130; Wibmer and O’Brien 1986a, 73, 1986b; Alonso-Zarazaga and Lyal 1999, 158; Morrone 1999, 123.

*Brachyomus quadrinodosus* (Boheman, 1842: 217)

Distribution: Colombia, Venezuela.

References: Boheman in Schönher 1842, 217 (as Geonemus), Wibmer and O’Brien 1986a, 74; Morrone 1999, 123.

*Brachyomus quadrituberculatus* (Boheman, 1842: 216)

Distribution: **Colombia**.

References: Boheman in Schönher 1842, 216 (as Geonemus), Wibmer and O’Brien 1986a, 74; Morrone 1999, 123.

Genus *Compsus* Schönher, 1823: col. 1140

Distribution: Argentina, Bolivia, Brazil, Colombia, Ecuador, French Guiana, Guyana, Peru, Surinam, Venezuela; Costa Rica, Guatemala, Mexico, Nicaragua, Panama; Guadeloupe, Jamaica; NC, SE, SW USA.


*Compsus adonis* Marshall, 1922: 197

Distribution: Colombia (Antioquia: **Medellín**).
References: Marshall 1922b, 197; Hustache 1938a, 71 (in key, as *Compsus bellus*, but see discussion in Girón and Chamorro 2020, 38), Wibmer and O’Brien 1986a, 74; Morrone 1999, 123, O’Brien and Peña 2012.

**Compsus aeruginosus** (Boheman, 1840: 177)

Distribution: Colombia (Antioquia).

References: Boheman in Schönherr 1840b, 177 (as *Platyomus aeruginosus*), Wibmer and O’Brien 1986a, 74; Morrone 1999, 123.

**Compsus affinis** Hustache, 1938: 77

Distribution: Colombia (Meta: Villavicencio).


**Compsus albosetosus** Hustache, 1938: 84

Distribution: Colombia (Cundinamarca: Bogotá).


**Compsus albus** Hustache, 1938: 76

Distribution: Colombia (Cundinamarca: Fusagasugá; Tolima: Ibagué).


**Compsus alternnevittatus** Hustache, 1938: 83

Distribution: Colombia (Rio Chili, Boyacá (Muzo), Pensilvania (Caldas (?), Cundinamarca (?))).


**Compsus ater** Kirsch, 1889: 20

Distribution: Colombia (Pasto, Santa Lucía).

Compsus attenuatus Hustache, 1938: 84

Distribution: Colombia (Rio Chili; Boyacá: Muzo; Quindío Tolima: Ibagué).

Compsus bicarinatus Kirsch, 1889: 22

Distribution: Colombia (Popayan: Silvia (?)).
References: Kirsch 1889, 22; Wibmer and O’Brien 1986a, 74; Morrone 1999, 124.

Compsus bituberculatus Kirsch, 1889: 19

Distribution: Colombia (Cauca [Popayan]: Puracé).
References: Kirsch 1889, 19; Wibmer and O’Brien 1986a, 74; Morrone 1999, 124.

Compsus bituberosus Kirsch, 1868: 237

Distribution: Colombia (‘Bogotá’).

Compsus candidus Hustache, 1938: 75

Distribution: Colombia (‘Bogotá’); Valle del Cauca: Cartago, Palmira.

Compsus canescens (Bohemian, 1840: 181)

Distribution: Colombia; Napo and Amazon rivers (Brazil (?), Ecuador (?), Peru (?)).
References: Bohemian in Schönherr 1840b, 181 as Platyomus canescens, Guérin-Méneville 1855, 592 (listed), Wibmer and O’Brien 1986a, 75; Morrone 1999, 124.
Compsus cyanitarsis Hustache, 1938: 93

Distribution: Bolivia, Colombia.

Compsus cyphoides Hustache, 1938: 108

Distribution: Colombia (Meta: Villavicencio).

Compsus delicatulus Hustache, 1938: 79

Distribution: Colombia (Valle del Cauca: Cartago).

Compsus deliciosus Hustache, 1938: 80

Distribution: Colombia (Risaralda: Pereira).

Compsus deplanatus Kirsch, 1868: 237

Distribution: Colombia (‘Bogotá’), Venezuela.
References: Kirsch 1868, 237; Wibmer and O’Brien 1986a, 75; Morrone 1999, 124.

Compsus divisus Hustache, 1938: 82

Distribution: Colombia (Boyacá: Muzo).

Compsus eustylodes Hustache, 1938: 96

Distribution: Colombia (Cundinamarca: Fusagasugá, Viotá; Meta: Villavicencio).

**Compsus glaucus** (Boheman, 1840: 178)

Distribution: Colombia (Antioquia).
References: Boheman in Schönherr 1840b, 178 as *Platyomus glaucus*, Wibmer and O’Brien 1986a, 75; Morrone 1999, 125.

**Compsus iris** Marshall, 1922: 198

Distribution: Colombia (Antioquia: Medellín).
References: Marshall 1922b, 198; Wibmer and O’Brien 1986a, 75; Morrone 1999, 125.

**Compsus latifrons** Hustache, 1938: 90

Distribution: Colombia (Pereira)

**Compsus lebasii** (Boheman, 1840: 182)

Distribution: Colombia (Bolívar: Cartagena).

**Compsus lineatus** Hustache, 1938: 81

Distribution: Colombia (S. Antonio)
References: Hustache 1938a, 81; Wibmer and O’Brien 1986a, 76; Morrone 1999, 125, O’Brien and Peña 2012, 3 (in key).

**Compsus obliquatus** Hustache, 1938: 77

Distribution: Colombia (Rio Chili; Cundinamarca: Beltrán; Tolima: Espinal).
Compsus pertinax Hustache, 1938: 85

Distribution: Colombia (Cauca), Ecuador.

Compsus placidus (Boheman, 1840: 180)

Distribution: Colombia [Nova Granata].
References: Boheman in Schönherr 1840b, 180 as Platyomus placidus, Wibmer and O’Brien 1986a, 76; Morrone 1999, 125.

Compsus popayanus Kirsch, 1889: 21

Distribution: Colombia (Popayán).
References: Kirsch 1889, 21; Wibmer and O’Brien 1986a, 76; Morrone 1999, 125.

Compsus pugionatus Marshall, 1922: 196

Distribution: Colombia, Venezuela; Central America.
References: Marshall 1922b, 196; Wibmer and O’Brien 1986a, 76; Morrone 1999, 125.

Compsus quadrisignatus (Boheman, 1840: 188)

Distribution: Colombia, French Guiana, Venezuela.
References: Boheman in Schönherr 1840b, 188 as Platyomus quadrisignatus, Wibmer and O’Brien 1986a, 76; Morrone 1999, 125.

Compsus roseomicans Hustache, 1938: 100

Distribution: Colombia (Boyacá: Muzo).
References: Hustache 1938a, 100; Wibmer and O’Brien 1986a, 76; Morrone 1999, 125, O’Brien and Peña 2012, 6 (in key).

Compsus scrutator Hustache, 1938: 98

Distribution: Brazil, Colombia (Cauca), Venezuela.
References: Hustache 1938a, 98; Wibmer and O’Brien 1986a, 76; Morrone 1999, 125, O’Brien and Peña 2012, 6 (in key).

**Compsus violaceus** Hustache, 1938: 89

Distribution: Colombia (Rio Chili).


**Compsus viridissimus** Hustache, 1938: 107

Distribution: Colombia (Boyacá: Muzo), Ecuador.


**Compsus viridivittatus** (Guérin-Méneville, 1855: 592)

Distribution: Colombia (Antioquia (Támesis), Caldas (Chinchiná), Cundinamarca (Beltrán), Quindío (Armenia, La Tebaida, Montenegro), Risaralda (Belén de Umbría, Pereira), Santander (Lebrija), Tolima (Armero Guayabal, 4 Km SE Ibagué), Valle del Cauca (Caicedonia, Sevilla); Napo and Amazon rivers (Brazil (?), Ecuador (?), Peru (?)).


**Compsus viridulus** Hustache, 1938: 95

Distribution: Colombia (‘Bogotá’).


**Compsus zebra** Marshall, 1922: 199

Distribution: Colombia (Valle del Cauca: Buenaventura), Ecuador.

**Compsus zebrinus** Voss, 1953: 57

Distribution: Colombia [West Cordilleran. Rio Aguacatal, St. Antonio, Monte Socorro].

References: Voss 1953, 57; Wibmer and O’Brien 1986a, 77; Morrone 1999, 126.

Genus *Eustylus* Schönherr, 1842: 40

Distribution: Bolivia, Brazil, Colombia, French Guiana, Guyana, Paraguay, Peru, Trinidad, Venezuela; Costa Rica, Guatemala, Mexico, Panama; Guadeloupe, St. Lucia.

References: Schönherr 1842, 40; Sharp and Champion 1911, 290; Marshall 1916 (most Colombian species), Wibmer and O’Brien 1986a, 73; Alonso-Zarazaga and Lyal 1999, 158; Morrone 1999, 127.

*Eustylus bodkini* Marshall, 1916: 456

Distribution: Colombia, Guyana, Venezuela.


Distribution: Colombia.


*Eustylus magdalenae* Marshall, 1926: 536

Distribution: Colombia (Magdalena: El Banco [El Blanco], Magdalena Valley; Aracataca).


*Eustylus puber* (Olivier, 1807: 367)

Distribution: Colombia, French Guiana, Guyana, Trinidad, Venezuela.

References: Olivier 1807, 367 (as *Curculio puber*), Wibmer and O’Brien 1986a, 73; Morrone 1999, 127; Franz 2010a, 56 (re-description).

*Eustylus simplex* Marshall, 1916: 460

Distribution: Colombia.

**Eustylus simulatus** Marshall, 1916: 459

Distribution: **Colombia**.


**Eustylus sordidus** Marshall, 1916: 460

Distribution: **Colombia**.


Genus *Exophthalmus* Schönherr, 1823: col. 1140

Distribution: Brazil, Colombia, Ecuador, French Guiana, Surinam; Belize, Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Cuba, Dominican Republic, Guadeloupe, Haiti, Jamaica, Martinique, Puerto Rico.


**Exophthalmus annulonotatus** (Waterhouse, 1879: 423)

Distribution: Colombia (Antioquia: Medellín), Ecuador.

References: Waterhouse 1879, 423 (as *Paepodes annulonotatus*), Wibmer and O’Brien 1986a, 79; Morrone 1999, 128.

**Exophthalmus consobrinus** (Marshall, 1922b: 190)

Distribution: Colombia (Chocó: Andagoya, Río Condoto; Valle del Cauca (Buenaventura).

References: Marshall 1922b, 190 (as *Exophthalmodes consobrinus*), Wibmer and O’Brien 1986a, 79; Morrone 1999, 128.

**Exophthalmus crassicornis** Kirsch, 1868: 238

Distribution: Colombia (‘Bogotá’); Panama.

**Exophthalmus jekelianus** (White, 1858: 357)

Distribution: Colombia (**Chocó**); Costa Rica, Nicaragua, Panama.

References: White in Jekel 1858, 357 (as *Praepodes jekelianus*), O’Brien and Wibmer 1982, 57; Wibmer and O’Brien 1986a, 79; Morrone 1999, 128.

**Exophthalmus sulcicrus** Champion, 1911: 268

Distribution: Colombia (**Chocó**); Costa Rica, Guatemala, Nicaragua, Panama.


**Genus** *Exorides* Pascoe, 1881: 43

Distribution: Colombia, Ecuador, Peru, Venezuela.

References: Pascoe 1881, 43; Marshall 1922b, 202; Wibmer and O’Brien 1986a, 80; Alonso-Zarazaga and Lyal 1999, 158; Morrone 1999, 130.

**Exorides bifurcatus** Marshall, 1926: 537

Distribution: Colombia (**Magdalena**: **Sierra de San Lorenzo**).


**Exorides caudatus** Marshall, 1922: 217

Distribution: Colombia (‘**Bogotá**’).


**Exorides cylindricus** Marshall, 1922: 213

Distribution: Colombia (**Magdalena, San Lorenzo Mt.**).

Exorides espeletiae (Kirsch, 1889: 23)

Distribution: Colombia (‘Bogotá’).
References: Kirsch 1889, 23 (as Compsus espeletiae), Marshall 1922b, 217; Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.

Exorides labyrinthicus (Kirsch, 1889: 22)

Distribution: Colombia (Pasto, Santa Lucía).
References: (Kirsch 1889, 22 (as Compsus labyrinthicus), Marshall 1922b, 203; Wibmer and O’Brien 1986a, 80; Morrone 1999, 130).

Exorides lindigi (Kirsch, 1889: 24)

Distribution: Colombia (Bogotá).
References: Kirsch 1889, 24 (as Compsus lindigi), Marshall 1922b, 205 (in key), Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.

Exorides marshalli (Bovie, 1908b: 44)

Distribution: Colombia.
References: Bovie 1908b, 44 (as Compsus marshalli), Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.

Exorides masoni Marshall, 1922: 215

Distribution: Colombia (Magdalena, San Lorenzo Mt.).

Exorides mucronatus (Faust, 1892: 18)

Distribution: Colombia, Venezuela.
References: Faust 1892, 18 (as Synthlibonotus mucronatus), Marshall 1922b, 214; Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.
Exorides obesus Marshall, 1922: 209

Distribution: Colombia.

Exorides pyriformis Marshall, 1922: 211

Distribution: Colombia.
  References: Marshall 1922b, 211; Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.

Exorides quadrivittatus (Kirsch, 1889: 23)

Distribution: Colombia (Pasto, Santa Lucía).
  References: Kirsch 1889, 23; Marshall 1922b, 206; Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.

Exorides rugosus (Taschenberg, 1870: 188)

Distribution: Colombia (‘Bogotá’).
  References: Taschenberg 1870, 188 (as Compsus rugosus), Marshall 1922b, 204; Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.

Exorides septemcostatus Marshall, 1922: 212

Distribution: Colombia, Ecuador.
  References: Marshall 1922b, 212; Wibmer and O’Brien 1986a, 80; Morrone 1999, 130.

Genus Oxyderces Schönherr, 1823: col. 1140

Distribution: Argentina, Bolivia, Brazil, Colombia, Ecuador, Paraguay, Peru, Venezuela; Guadeloupe, Martinique.
**Oxyderces exaratus** (Hustache, 1938a: 111)

Distribution: Colombia [Columbia] ([Transylvanie] Pensilvania, Cundinamarca (?)).

References: Hustache 1938a, 111 (as Plococompsus exaratus), Wibmer and O’Brien 1986a, 81; Morrone 1999, 130.

**Oxyderces mirandus** (Pascoe, 1880: 423)

Distribution: Colombia [Columbia] (Boyacá: Muzo), Ecuador.

References: Pascoe 1880, 423; Marshall 1922b, 201 (listed as Compsus mirandus), Hustache 1938a, 114 (as Plococompsus texatus), Wibmer and O’Brien 1986a, 81; Morrone 1999, 131.

**Oxyderces viridiaeris** (Hustache, 1938a: 115)

Distribution: Colombia.

References: Hustache 1938a, 115 (as Plococompsus viridiaeris), Wibmer and O’Brien 1986a, 81; Morrone 1999, 131.

**Oxyderces viridipes** (Boheman, 1840: 179)

Distribution: Colombia (Antioquia).

References: Boheman in Schönherr 1840b, 179 (as Platyomus viridipes), Wibmer and O’Brien 1986a, 81; Morrone 1999, 131.

Genus *Synthlibonotus* Schönherr, 1847: 41 [91]

Distribution: Colombia, Venezuela; Guatemala, Mexico.


*Synthlibonotus rufipes* Schönherr, 1847: 41 [91]

Distribution: Colombia, Venezuela.

References: Schönherr 1847, 41 [91], Wibmer and O’Brien 1986a, 81; Morrone 1999, 132.
Genus *Xestogaster* Marshall, 1922: 221

Distribution: Brazil, Colombia, Peru.

**Xestogaster porosa** Marshall, 1922: 221

Distribution: Colombia (‘Bogotá’).
References: Marshall 1922b, 221; Wibmer and O'Brien 1986a, 81; Morrone 1999, 132.

**Xestogaster squalida** Marshall, 1922: 222

Distribution: Colombia.

**Tribe Lordopini Schönherr, 1823: col. 1142**

Genus *Acanthobrachis* Jekel, 1854 [1857]-I: 9bis (in key) [1857: 101]

Distribution: Brazil, Colombia.

*Acanthobrachis germari* Jekel, 1857: 103

Distribution: Brazil, Colombia.

**Genus Atomorhinus Hustache, 1946: 3**

Distribution: Colombia.
**Atomorhinus impressidorsum** Hustache, 1946: 4

Distribution: Colombia (Cauca, Rio Chili; Cundinamarca: Fusagasugá).

Genus **Deroconus** Jekel, 1854 [1857]-I: 9bis (in key)

Distribution: Colombia, Venezuela.

*Deroconus rufipes* (Lacordaire, 1863: 269)

Distribution: **Colombia**, Venezuela.
References: Lacordaire 1863, 269 (as *Hypsonotus rufipes*), Kessel 1932, 66 (as *Deraconus rufipes*), Wibmer and O’Brien 1986a, 93; Morrone 1999, 111.

**Genus Eudmetus** Jekel, 1856: 9bis [1857: 105]

Distribution: Colombia.

**Eudmetus cinereus** Jekel, 1857: 107

Distribution: **Colombia**.

**Eudmetus nigromaculatus** Jekel, 1857: 111

Distribution: **Colombia**.
References: Jekel 1856, 111; Wibmer and O’Brien 1986a, 91; Morrone 1999, 112.

**Eudmetus posticatus** Jekel, 1857: 109

Distribution: **Colombia**.
Genus *Granadia* Kessel, 1935: 95

Distribution: Colombia.

*Granadia humeralis* Kessel, 1935: 96

Distribution: Colombia.

Genus *Hypoptophila* Voss, 1934: 102

Distribution: Brazil, Colombia.

*Hypoptophila munda* Voss, 1934: 102

Distribution: Colombia [Nov. Granada].

Genus *Hypoptus* Jekel 1856: 9bis (in key)

Distribution: Bolivia, Colombia, Ecuador; Belize, Costa Rica, El Salvador, Guatemala, Mexico, Nicaragua, Panama; Grenada, St. Vincent.

*Hypoptus arcticus* Kessel, 1932: 62


*Hypoptus lepyroides* Kessel, 1932: 63

Distribution: Colombia [Columbien].
References: Kessel 1932, 63; Wibmer and O’Brien 1986a, 93; Morrone 1999, 113.

*Hypoptus macularis* Champion, 1911: 303

Distribution: Colombia; Belize, Costa Rica, El Salvador, Guatemala, Mexico, Nicaragua, Panama.


*Hypoptus setosulus* Kessel, 1932: 64

Distribution: **Colombia**.

References: Kessel 1932, 64; Wibmer and O’Brien 1986a, 93; Morrone 1999, 113.

Genus *Hypsonotus* Germar, 1824: 367

Distribution: Argentina, Bolivia, Brazil, Colombia, Ecuador, Paraguay, Venezuela; Costa Rica, Guatemala, Honduras, Mexico; St. Vincent.


*Hypsonotus acutipennis* Jekel, 1859: 229

Distribution: Colombia (‘Bogotá’).

References: Jekel 1856, 160 (listed), 1859, 229 (described), Wibmer and O’Brien 1986a, 97; Morrone 1999, 113.

*Hypsonotus apicatus* Jekel, 1857: 173

Distribution: Colombia (‘Bogotá’).

References: Jekel 1857, 145 (listed), 1859, 173 (described), Wibmer and O’Brien 1986a, 97; Morrone 1999, 114.

*Hypsonotus bipunctatus* Jekel, 1859: 233

Distribution: **Colombia**.
References: Jekel 1856, 160 (listed), 1859, 233 (described), Wibmer and O’Brien 1986a, 97; Morrone 1999, 114.

Hypsonotus callosicollis Jekel, 1857: 179

Distribution: Brazil, Colombia.

References: Jekel 1857, 146 (listed), 179 (described), Wibmer and O’Brien 1986a, 97; Morrone 1999, 114.

Hypsonotus compressipennis Jekel, 1859: 235

Distribution: Colombia.

References: Jekel 1857, 160 (listed), 1859, 235 (described), Wibmer and O’Brien 1986a, 97; Morrone 1999, 114.

Hypsonotus glaber Kessel, 1937: 170

Distribution: Colombia.


Hypsonotus hondurensis Kessel, 1937: 161

Distribution: Colombia; Honduras.


Hypsonotus interior Kessel, 1937: 171

Distribution: Colombia.


Hypsonotus laevicollis Jekel, 1859: 227

Distribution: Colombia (‘Bogotá’).
References: Jekel 1857, 160 (listed), 1859, 227 (described), Wibmer and O’Brien 1986a, 98; Morrone 1999, 115

**Hypsonotus latissimus** Kessel, 1937: 125

Distribution: Colombia.

References: Kessel 1937, 125 (not seen), Wibmer and O’Brien 1986a, 98; Morrone 1999, 115.

**Hypsonotus limbifer** Kessel, 1937: 155

Distribution: Colombia.


**Hypsonotus nitidulus** Jekel, 1859: 237

Distribution: Colombia.


**Hypsonotus obsoletus** Jekel, 1857: 155

Distribution: Brazil, Colombia.

References: Jekel 1857, 155 (listed), 1859, 227 (described), Wibmer and O’Brien 1986a, 99; Morrone 1999, 115.

**Hypsonotus punctum** Jekel, 1859: 231

Distribution: Colombia.


**Hypsonotus ramosus** Jekel, 1859: 225

Distribution: Colombia.

**Hypsonotus setarius** Jekel, 1857: 156

Distribution: **Colombia**.


**Hypsonotus vestitus** Jekel, 1859: 223

Distribution: **Colombia**.


Genus **Lordops** Schön herr, 1823: col. 1142

Distribution: Argentina, Brazil, Colombia, Paraguay, Venezuela; Mexico.


**Lordops conjugatus** Jekel, 1854: I.V.7

Distribution: Brazil, Colombia (‘Bogotá’), Paraguay.

References: Jekel 1854, I.V.7, Kessel 1932, 7 (in key; p. 30 comments), Wibmer and O’Brien 1986a, 90; Morrone 1999, 117.

Genus **Orthocnemus** Jekel, 1856: 131

Distribution: Colombia, Ecuador, Peru.

References: Jekel 1856, 131; Kessel 1932, 40; Wibmer and O’Brien 1986a, 92; Alonso-Zarazaga and Lyal 1999, 162; Morrone 1999, 118.

**Orthocnemus lebasii** Jekel, 1857: 133

Distribution: Colombia (‘Bogotá’).

References: Jekel 1857, 133; Kessel 1932, 41; Wibmer and O’Brien 1986a, 92; Morrone 1999, 118.
Genus *Sulla* Kessel, 1937: 182

Distribution: Colombia.
References: Kessel 1937, 182; Wibmer and O’Brien 1986a, 91 (as *Sullana*), Alonso-Zarazaga and Lyal 1999, 162; Morrone 1999, 121 (as *Sullana*).

*Sulla columbiana* Kessel, 1937: 183

Distribution: Colombia.
References: Kessel 1937, 183; Wibmer and O’Brien 1986a, 91 (as *Sullana columbiana*), Morrone 1999, 121 (as *Sullana columbiana*).

Tribe Naupactini Gistel, 1856: 374

Genus *Amphideritus* Schönherr, 1840: 117

Distribution: Bolivia, Chile, Colombia, Peru.

*Amphideritus rugicollis* Kirsch, 1868: 223

Distribution: Colombia (‘Bogotá’).
References: Kirsch 1868, 223; Wibmer and O’Brien 1986a, 55; Morrone 1999, 156.

*Amphideritus setosus* (Schönherr, 1847: 29)

Distribution: *Colombia* (Bogotá).
References: Schönherr 1847, 29 (as *Pterotropis setosus*); Kirsch 1868, 224 (as *Amphideritus squamosus*, a synonym), Wibmer and O’Brien 1986a, 55; Morrone 1999, 156.

*Amphideritus vilis* Boheman, 1840: 118

Distribution: Colombia (Bogotá).
References: Boheman in Schönherr 1840b, 118; Wibmer and O’Brien 1986a, 55; Morrone 1999, 156.

Distribution: Colombia, Ecuador.


*Asymmathetes steinheili* (Kirsch, 1889: 13)

Distribution: Colombia (*Consacá*).

References: Kirsch 1889, 13 (as *Naupactus steinheili*), Wibmer and O’Brien 1986a, 53; Morrone 1999, 151.

Genus *Chamaelops* Kirsch, 1868: 235

Distribution: Colombia.


*Chamaelops fissirostris* (Chevrolat, 1878: LV)

Distribution: Colombia (*Bogotá*).

References: Chevrolat 1878, LV (as *Temnoscapus fissirostris*), Wibmer and O’Brien 1986a, 51; Morrone 1999, 151.

*Chamaelops munitus* Kirsch, 1868: 236

Distribution: Colombia (*Bogotá*).

References: Kirsch 1868, 236; Wibmer and O’Brien 1986a, 51; Morrone 1999, 152.

Genus *Ericydeus* Pascoe, 1880: 422

Distribution: Argentina, Bolivia, Brazil, Colombia, Ecuador, French Guiana, Peru, Venezuela; Costa Rica, Mexico; SW USA.

*Ericydeus nigropunctatus* (Chevrolat, 1877: 170)

Distribution: Colombia (Guainía), Ecuador, Peru, Venezuela.

References: Chevrolat 1877, 170 (as *Cyphus nigropunctatus*), Wibmer and O’Brien 1986a, 51; Lanteri 1995, 402; Morrone 1999, 152.

*Ericydeus sedecimpunctatus* (Linnaeus, 1758: 386)

Distribution: Argentina, Bolivia, Brazil, Colombia, French Guiana, Guyana, Venezuela; Costa Rica, Panama.


Genus *Hoplopactus* Jekel, 1875: 138

Distribution: Bolivia, Brazil, Colombia, Ecuador, French Guiana, Peru, Trinidad, Venezuela.


*Hoplopactus dentipes* (Kirsch, 1868: 231)

Distribution: Colombia (‘Bogotá’).


*Hoplopactus pavidus* (Boheman, 1840: 107)

Distribution: Colombia, Trinidad, Venezuela.


*Hoplopactus rufipes* (Kirsch, 1868: 230)

Distribution: Colombia (‘Bogotá’).

**Hoplopatcus suturalis** (Kirsch, 1868: 230)

Distribution: Colombia (‘Bogotá’).

Genus *Lanterius* Alonso-Zarazaga y Lyal, 1999

Distribution: Argentina, Brazil, Colombia, Ecuador, Paraguay, Venezuela; Mexico. Panama.

*Lanterius amandus* (Kirsch, 1868: 225)

Distribution: Colombia (‘Bogotá’); Panama.
References: Kirsch 1868, 225 (as *Mimographus amandus*), Wibmer and O’Brien 1986a, 56 (as *Mimographus amandus*), Alonso-Zarazaga and Lyal 1999, 164; Morrone 1999, 154 (as *Macrostyles amandus*).

*Lanterius aridosiacus* (Kirsch, 1868: 229), *comb. nov.*

Distribution: Colombia (‘Bogotá’).
References: Kirsch 1868, 229 (as *Mimographus aridosiacus*), Wibmer and O’Brien 1986a, 56 (as *Mimographus aridosiacus*), Morrone 1999, 155 (as *Macrostyles aridosiacus*).

*Lanterius cinereoguttatus* (Champion, 1911: 228), *comb. nov.*

Distribution: Colombia, Venezuela; Central America (unspecified).
References: Sharp and Champion 1911, 228 (as *Steirarrhinus cinereoguttatus*), Wibmer and O’Brien 1986a, 56 (as *Mimographus cinereoguttatus*), Morrone 1999, 155 (as *Macrostyles cinereoguttatus*).

*Lanterius hirtus* (Voss, 1953: 60), *comb. nov.*

Distribution: Colombia ([West Cordilleren, Rio Aguacatal]).
References: Voss 1953, 60 (as *Neoanypotactus hirtus*), Wibmer and O’Brien 1986a, 56 (as *Mimographus hirtus*), Morrone 1999, 155 (as *Macrostyles hirtus*).
Lanterius laesicollis (Schönherr, 1847: 35), *comb. nov.*

Distribution: Colombia, Venezuela.
References: Schönherr 1847, 35 (as *Mimographus laesicollis*), Wibmer and O’Brien 1986a, 56 (as *Mimographus laesicollis*), Morrone 1999, 155 (as *Macro stylus laesicollis*).

*Lanterius micans* (Kirsch, 1868: 229), *comb. nov.*

Distribution: Colombia (‘Bogotá’).
References: Kirsch 1868, 229 (as *Mimographus micans*), Wibmer and O’Brien 1986a, 56 (as *Mimographus micans*), Morrone 1999, 155 (as *Macro stylus micans*).

*Lanterius rotundicollis* (Kirsch, 1889: 15), *comb. nov.*

Distribution: Colombia (Popayán).
References: Kirsch 1889, 15 (as *Mimographus rotundicollis*), Wibmer and O’Brien 1986a, 56 (as *Mimographus rotundicollis*), Morrone 1999, 156 (as *Macro stylus rotundicollis*).

*Lanterius versicolor* (Kirsch, 1889: 15), *comb. nov.*

Distribution: Colombia (‘Pasto’).
References: Kirsch 1889, 15 (as *Mimographus versicolor*), Wibmer and O’Brien 1986a, 56 (as *Mimographus versicolor*), Morrone 1999, 156 (as *Macro stylus versicolor*).

*Lanterius vittatus* (Kirsch, 1889: 14), *comb. nov.*

Distribution: Colombia (‘Pasto’).
References: Kirsch 1889, 14 (as *Mimographus vittatus*), Wibmer and O’Brien 1986a, 56 (as *Mimographus vittatus*), Morrone 1999, 156 (as *Macro stylus vittatus*).

Genus *Leschenius* del Río, Marvaldi y Lanteri, 2012: 55

Distribution: Colombia, Ecuador.
References: del Río et al. 2012, 55.
Leschenius vulcanorum (Kirsch, 1889: 17)

Distribution: Colombia (Nariño), Ecuador.

   References: Kirsch 1889, 17 (as Canephorus vulcanorum), Wibmer and O’Brien 1986a, 53 (as Asymmathetes vulcanorum), Canchala 1992; Morrone 1999, 151 (as Asymmathetes vulcanorum), del Río et al. 2012, 62.

Genus Litostylus Faust, 1894: 368

Distribution: Argentina, Brazil, Colombia, French Guiana, Peru, Venezuela; Honduras, Nicaragua, Panama; Antigua, Barbados, Dominica, Guadeloupe, Martinique, Montserrat, Saint Barthelemy, Saint Vincent.


Litostylus diadema (Fabricius, 1787: 116)

Distribution: Argentina, Brazil, Colombia, French Guiana, Peru, Venezuela; Honduras, Nicaragua, Panama.


Genus Melanocyphus Jekel, 1875: 143

Distribution: Colombia.


Melanocyphus bispinus (Boheman, 1840: 150)

Distribution: Colombia (Bogotá).

   References: Boheman in Schönherr 1840b, 150 (as Cyphus bispinus), Wibmer and O’Brien 1986a, 54; Morrone 1999, 156, del Río and Lanteri 2007a, 131.

Melanocyphus lugubris (Boheman, 1840: 147)

Distribution: Colombia (Bogotá).

   References: Boheman in Schönherr 1840b, 147 (as Naupactus lugubris), Wibmer and O’Brien 1986a, 54; Morrone 1999, 156, del Río and Lanteri 2007a, 130.
Genus *Mimographus* Schönherr, 1847: 34

Distribution: Colombia, Peru; Belize, Costa Rica, Guatemala, Mexico, Nicaragua, Panama.

References: Schönherr 1847, 34; Wibmer and O’Brien 1986a, 56 (as *Steirarrhinus*), Alonso-Zarazaga and Lyal 1999, 165; Morrone 1999, 154 (as *Macrostylus*).

*Mimographus decolor* (Boheman, 1840: 106)

Distribution: Colombia; Central America (unspecified).

References: Boheman in Schönherr 1840b, 106 (as *Naupactus decolor*), Sharp and Champion 1911, 223 (as *Steirarrhinus conicollis*, a synonym), O’Brien and Wibmer 1982, 32 (as *Steirarrhinus conicollis*), Wibmer and O’Brien 1986a, 56 (as *Steirarrhinus decolor*), Morrone 1999, 155 (as *Macrostylus decolor*).

*Mimographus depressicollis* (Boheman, 1840: 66)

Distribution: Colombia (Antioquia).

References: Boheman in Schönherr 1840b, 66 (as *Naupactus depressicollis*), Wibmer and O’Brien 1986a, 56 (as *Steirarrhinus depressicollis*), Morrone 1999, 155 (as *Macrostylus depressicollis*).

*Mimographus elegantulus* (Boheman, 1840: 64)

Distribution: Colombia (Antioquia).

References: Boheman in Schönherr 1840b, 64 (as *Naupactus elegantulus*), Wibmer and O’Brien 1986a, 56 (as *Steirarrhinus elegantulus*), Morrone 1999, 155 (as *Macrostylus elegantulus*).

*Mimographus plicaticollis* (Hustache, 1938b: 274)

Distribution: Colombia (Cundinamarca (Fusagasugá), Valle del Cauca (Cali), San Antonio, Rio Cauca).

References: Hustache 1938b, 274 (as *Naupactus plicaticollis*), Wibmer and O’Brien 1986a, 56 (as *Steirarrhinus plicaticollis*), Morrone 1999, 155 (as *Macrostylus plicaticollis*).

*Mimographus varians* (Boheman, 1840: 65)

Distribution: Colombia (Antioquia).
References: Boheman in Schönherr 1840b, 65 (as Naupactus varians), Wibmer and O’Brien 1986a, 56 (as Steirarrhinus varians), Morrone 1999, 155 (as Macrostylus varians).

Genus Naupactus Dejean, 1821: 94

Distribution: Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, French Guiana, Guyana, Paraguay, Peru, Surinam, Uruguay, Venezuela; Costa Rica, Guatemala, Honduras, Mexico; SE, SW USA.


Naupactus instabilis Boheman, 1840: 71

Distribution: Colombia, Venezuela.

References: Boheman in Schönherr 1840b, 71; Wibmer and O’Brien 1986a, 60; Morrone 1999, 159.

Naupactus venezolanus Hustache, 1938: 270

Distribution: Colombia, Venezuela.


Genus Platyomus Sahlberg, 1823: 29

Distribution: Argentina, Bolivia, Brazil, Colombia**, Ecuador, Guyana, Paraguay, Peru, Uruguay, Venezuela; Belize, Costa Rica, El Salvador, Guatemala, Honduras. Mexico; SW USA (Texas).


Undetermined species

Distribution: Colombia (Amazonas, Antioquia, Meta, Quindío, Risaralda, Valle del Cauca).

References: Girón 2007b; Cardona-Duque et al. 2018, 254–255.
Genus *Plectrophoroides* Wibmer and O’Brien, 1986: 50

Distribution: Argentina, Bolivia, Brazil, Colombia, French Guiana, Guyana, Peru, Surinam, Venezuela.

*Plectrophoroides acuminatus* (Chevrolat, 1879: LXXVII)

Distribution: Colombia.
References: Chevrolat 1879, LXXVII (as *Plectophorus acuminatus*), Wibmer and O’Brien 1986a, 50; Morrone 1999, 164.

*Plectrophoroides albilabris* (Chevrolat, 1879: LXXVIII)

Distribution: Colombia (’Bogotá’).
References: Chevrolat 1879, LXXVIII (as *Plectophorus albilabris*), Wibmer and O’Brien 1986a, 50; Morrone 1999, 164.

*Plectrophoroides bifasciatus* (Chevrolat, 1879: LXXVII)

Distribution: Bolivia, Colombia.
References: Chevrolat 1879, LXXVII (as *Plectophorus bifasciatus*), Wibmer and O’Brien 1986a, 50; Morrone 1999, 164.

Genus *Tetragononomus* Champion, 1911: 240

Distribution: Colombia; Panama.

*Tetragononomus tuberosus* Champion, 1911: 240

Distribution: Colombia; Panama.
Tribe Premnotrypini Kuschel, 1956: 187

Genus *Premnotrypes* Pierce, 1914: 348

Distribution: Bolivia, Chile, Colombia, Ecuador, Peru, Venezuela.

*Premnotrypes vorax* (Hustache, 1933: 377)

Distribution: Colombia (‘Bogotá’).

Tribe Tanymecini Lacordaire, 1863: 82

Genus *Airosimus* Howden, 1966: 174

Distribution: Bolivia, Brazil, Colombia, Ecuador, Peru, Paraguay, Venezuela; Costa Rica, Mexico, Panama.

*Airosimus robustus* (Faust, 1892: 4)

Distribution: Colombia, Venezuela.
References: Faust 1892, 4 (as *Menetypus robustus*), Howden 1966, 196; Wibmer and O’Brien 1986a, 71; Morrone 1999, 143.

*Airosimus semirobustus* Howden, 1966: 199

Distribution: Colombia.
References: Howden 1966, 199; Wibmer and O’Brien 1986a, 71; Morrone 1999, 143.

Genus *Hadromeropsis* Pierce, 1913: 400

Distribution: Argentina, Bolivia, Brazil, Colombia, Ecuador, Paraguay, Peru, Uruguay, Venezuela; Costa Rica, Guatemala, Mexico, Panama.
Broad-nosed weevils of Colombia (Coleoptera, Curculionidae, Entiminae)

References: Pierce 1913, 400; Howden 1982; Wibmer and O’Brien 1986a, 68; Alonso-Zarazaga and Lyal 1999, 179; Morrone 1999, 143.

**Hadromeropsis alacer** Howden, 1982: 85

Distribution: **Colombia** (Bogotá, Valle del Cauca), Ecuador.
References: Howden 1982, 85; Wibmer and O’Brien 1986a, 68; Morrone 1999, 143.

**Hadromeropsis annae** Anderson, 2008: 65

Distribution: Colombia (**Páramo Sumapaz**).

**Hadromeropsis gemmifera** (Boheman, 1845: 418)

Distribution: Colombia (Bogotá, Magdalena, Santander), **Venezuela**; Guatemala, Panama.
References: Boheman in Schönherr 1845, 418 (as *Hadromerus gemmifer*), Howden 1982, 51 (as *Hadromeropsis gemmifer*), Wibmer and O’Brien 1986a, 69; Morrone 1999, 144.

**Hadromeropsis impressicollis** (Kirsch, 1868: 233)

References: Kirsch 1868, 223 (as *Hadromerus impressicollis*), Howden 1982, 94; Wibmer and O’Brien 1986a, 69; Morrone 1999, 144.

**Hadromeropsis magica** (Pascoe, 1881: 41)

Distribution: Brazil (?), Colombia (Bogotá, Cundinamarca (Fusagasugá)).
References: Pascoe 1881, 41 (as *Naupactus magicus*), Howden 1982, 114 (as *Hadromeropsis magicus*), Wibmer and O’Brien 1986a, 69; Morrone 1999, 144.

**Hadromeropsis mandibularis** Howden, 1982: 117

Distribution: Colombia (**St. Antonio** (prob. Valle del Cauca)).

*Hadromeropsis meridiana* Howden, 1982: 54

Distribution: Brazil, Colombia (Cundinamarca (Monte Redondo nr. Bogotá, Bogotá, nr. Girardot, Quetame), Tolima (Ibagué)).


*Hadromeropsis nebulicola* Howden, 1982: 88

Distribution: Colombia (Magdalena (San Lorenzo)).


*Hadromeropsis pectinata* Howden, 1982: 108

Distribution: Bolivia, Colombia, Peru.


*Hadromeropsis silacea* Howden, 1982: 90

Distribution: Brazil (?), Colombia (Antioquia (Frontino), Cauca).

References: Howden 1982, 90 (as *Hadromeropsis silaceus*), Wibmer and O’Brien 1986a, 69; Morrone 1999, 144.

*Hadromeropsis striata* Howden, 1982: 126

Distribution: Colombia [Paso Bella Vista, above Duriamaina].

References: Howden 1982, 126 (as *Hadromeropsis striatus*), Wibmer and O’Brien 1986a, 69; Morrone 1999, 144.

Genus *Macropterus* Schönherr, 1840: 419

Distribution: Brazil, Colombia, Peru.
References: Schönherr 1840b, 419; Wibmer and O’Brien 1986a, 72; Alonso-Zarazaga and Lyal 1999, 179; Morrone 1999, 145.

*Macropterus chlorostomus* Boheman, 1840: 424

Distribution: Brazil, Colombia.

References: Boheman in Schönherr 1840b, 424; Wibmer and O’Brien 1986a, 72; Morrone 1999, 145.

Genus *Pandeleteius* Schönherr, 1834: 129

Distribution: Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador, Paraguay, Peru, Venezuela; Costa Rica, El Salvador, Guatemala, Honduras, Mexico, Nicaragua, Panama; Dominica, Grenada, Guadeloupe, Jamaica, Saint Vincent; E Canada, USA.


*Pandeleteius admirabilis* Howden, 1976: 192

Distribution: Colombia (*Cauca*).


*Pandeleteius andeanus* Howden, 1976: 155

Distribution: Colombia (Boyacá (*Guateque*), Cundinamarca (Guasca-Guachetá, Mesitas del Colegio), Norte de Santander (*Chinácota*, Pamplona), Valle del Cauca (*Palmira*)), Venezuela.


*Pandeleteius antiochensis* Howden, 1976: 37

Distribution: Colombia (Antioquia (*Sonsón*)).

References: Howden 1976, 37; Wibmer and O’Brien 1986a, 70; Morrone 1999, 146.

*Pandeleteius arcanus* Howden, 1876: 60

Distribution: Colombia (*?*), Venezuela.

*Pandeleteius bordoni* Howden, 1976: 171

Distribution: Colombia (Norte de Santander (*Pamplona*)).
References: Howden 1976, 171; Wibmer and O’Brien 1986a, 70; Morrone 1999, 146.

*Pandeleteius campbelli* Howden, 1976: 104

Distribution: Colombia (Magdalena (*San Lorenzo*)).
References: Howden 1976, 104; Wibmer and O’Brien 1986a, 70; Morrone 1999, 146.

*Pandeleteius campestris* Howden, 1976: 73

Distribution: Colombia (Atlántico (Puerto Colombia), Bolívar (Cartagena, Turbaco), Cesar (Valledupar), Córdoba (Lorica), Magdalena (*Rio Frío*, Sevilla), Sucre (Toluviejo)), Venezuela.

*Pandeleteius carinipenis* Howden, 1976: 123

Distribution: Colombia (Valle del Cauca (*Saladito*)).
References: Howden 1976, 123; Wibmer and O’Brien 1986a, 70; Morrone 1999, 146.

*Pandeleteius chapini* Howden, 1976: 68

Distribution: Colombia (Meta (Restrepo, Villavicencio)), Venezuela.

*Pandeleteius clivus* Howden, 1976: 146

Distribution: Colombia (Cauca (*Silvia*)).
References: Howden 1976, 146; Wibmer and O’Brien 1986a, 70; Morrone 1999, 146.
**Pandeleteius conirostris** Howden, 1976: 101

Distribution: Colombia (Magdalena (San Lorenzo)).
References: Howden 1976, 101; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147.

**Pandeleteius dissimilis** Voss, 1939: 337

Distribution: Colombia (Candelaria (Valle del Cauca (?))); Costa Rica, Honduras, Panama.
References: Voss 1939, 337; Howden 1976, 185; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147.

**Pandeleteius eberhardi** Howden, 1976: 135

Distribution: Colombia (Valle del Cauca: Pichindé).
References: Howden 1976, 135; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147.

**Pandeleteius excisus** Howden, 1976: 169

Distribution: Colombia (Cundinamarca (La Aguadita, Fusagasugá, Silvania, Tequendama)).
References: Howden 1976, 169; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147.

**Pandeleteius flavus** Howden, 1976: 25

Distribution: Colombia (Valle del Cauca (Cali (Hacienda Arizona, Río Jamundí), Río Pance)).

**Pandeleteius giganteus** Howden, 1976: 94

Distribution: Colombia (Cauca (Silvia)).
References: Howden 1976, 94; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147.
**Pandeleteius hadromeroides** (Kirsch, 1868: 234)

Distribution: Colombia (‘Bogotá’, Cundinamarca (Bogotá, Anolaima, Fusagasugá, Guayabetal, La Aguadita, La Vega, Monte Redondo, Las Tibayes (between Honda and Bogotá), Tena), Quindío (Calarca)), Venezuela.

References: Kirsch 1868, 234 (as *Menetypus hadromeroides*), Howden 1976, 149; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147; Solano Rojas and Girón 2019.

**Pandeleteius hercules** Howden, 1976: 109

Distribution: Colombia (Magdalena (San Lorenzo)), Venezuela.

References: Howden 1976, 109; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147.

**Pandeleteius humboldti** Howden, 1976: 45

Distribution: Colombia (Cundinamarca (Tequendama, Mosquera, Zipaquirá-Pacho)).

References: Howden 1976, 45; Wibmer and O’Brien 1986a, 70; Morrone 1999, 147.

**Pandeleteius minax** Dohrn, 1880: 157

Distribution: Colombia (‘Bogotá’).


**Pandeleteius mirirostris** Howden, 1976: 23

Distribution: Colombia (Cundinamarca (Bogotá, Fusagasugá, Melgar, Mesitas del Colegio)).


**Pandeleteius modestus** (Faust, 1892: 3)

Distribution: Colombia (Norte de Santander (Santiago)), Venezuela.

Pandeleteius naupactoides (Pascoe, 1881: 38)

Distribution: Brazil, Colombia (Cundinamarca).

Pandeleteius nodifer Champion, 1911: 206

Distribution: Colombia (Bolívar (Cartagena), Cesar (Valledupar), Magdalena (Río Frío, Santa Marta, Sevilla)), Venezuela; Nicaragua, El Salvador; Honduras; Jamaica; SE SW USA.

Pandeleteius notabilis Howden, 1976: 128

Distribution: Colombia (Cauca (Silvia), Galego (Department unknown)).

Pandeleteius novagranadae Howden, 1976: 80

Distribution: Colombia (Norte de Santander (Chinácota, Pamplona)), Venezuela.

Pandeleteius olympus Howden, 1976: 57

Distribution: Colombia (Cauca (Silvia)).

Pandeleteius peckorum Howden, 1976: 125

Distribution: Colombia (Norte de Santander (Chinácota), Venezuela.

Pandeleteius pilosipectus Howden, 1976: 159

Distribution: Colombia (Cundinamarca (Bogotá, Laguna Ubaque)), Ecuador.
Pandeleteius procollis Howden, 1976: 84

Distribution: Colombia (Norte de Santander (Quebrada Honda)), Venezuela.
References: Howden 1976, 84; Wibmer and O’Brien 1986a, 71; Morrone 1999, 148.

Pandeleteius pygmaeus Howden, 1976: 70

Distribution: Colombia (Quindío (Calarcá), Valle del Cauca (Cali (Hacienda La Arizona, Río Jamundí, Río Pance, nr. Pinchindé)).

Pandeleteius reductus Howden, 1976: 141

Distribution: Colombia (Antioquia (El Retiro)).

Pandeleteius regina Howden, 1976: 177

Distribution: Colombia (Cundinamarca (Bogotá, La Unión-Bogotá), Norte de Santander (Pamplona)), Venezuela.

Pandeleteius santamartae Howden, 1976: 165

Distribution: Colombia (Magdalena (San Lorenzo)).

Pandeleteius scutellatus Howden, 1976: 97

Distribution: Colombia (Cundinamarca (Bogotá)).

Pandeleteius separatus Howden, 1976: 51

Distribution: Colombia (Norte de Santander (Pamplona)).
Pandeleteius subtilis Howden, 1996: 887

Distribution: Colombia (Meta (Restrepo)), Peru, Venezuela.

Pandeleteius summus Howden, 1976: 143

Distribution: Colombia (Cauca (Silvia)).

Pandeleteius tessellatus Howden, 1976: 189

Distribution: Colombia.
References: Howden 1976, 189; Wibmer and O’Brien 1986a, 71; Morrone 1999, 149.

Pandeleteius tinctorius Howden, 1976: 116

Distribution: Colombia (Cundinamarca (Fusagasugá, Zipaquirá-Pacho)).
References: Howden 1976, 116; Wibmer and O’Brien 1986a, 71; Morrone 1999, 149.

Pandeleteius torquatus Howden, 1976: 62

Distribution: Colombia (Cauca (Silvia)).
References: (Howden 1976, 62; Wibmer and O’Brien 1986a, 71; Morrone 1999, 149.

Pandeleteius truncatus Howden, 1976: 138

Distribution: Colombia (Cauca (Silvia), Valle del Cauca (Cali)).

Pandeleteius upsilon Howden, 1976: 90

Distribution: Colombia (Antioquia (Rionegro), Valle del Cauca (Lago Calima)).
References: Howden 1976, 90; Wibmer and O’Brien 1986a, 71; Morrone 1999, 149.
**Pandeleteius vitticollis** Champion, 1911: 202

Distribution: Colombia (Cundinamarca (Arbeláez, Bogotá, Fusagasugá)); Mexico, Guatemala, Honduras, Nicaragua, Panama.

References: Sharp and Champion 1911, 202; Howden 1976, 162; Wibmer and O’Brien 1986a, 71; Morrone 1999, 149.

**Additional comments**

This paper, as a compilation of information, constitutes the first step towards a better understanding of the biodiversity of entimines as a whole in Colombia and northern South America. It also highlights the areas and groups where a lot of work has been done and groups in need of revisionary work.

Due to its geographic location and attributes, Colombia harbours elements of different faunal components of the Americas: Andean, Amazon, Caribbean, Pacific and Orinoco (Ministerio del Medio Ambiente, Departamento Nacional de Planeación, Instituto Alexander von Humboldt 1996). Colombia is in a key position to contribute to the understanding of distributional, ecological and evolutionary patterns of Entiminae across the Americas, but remains as one of the most prominent knowledge gaps in the region for this subfamily and plenty of other insect taxa.

Except for economically-important species, there is a general lack of knowledge about the natural history of Colombian entimines (e.g. host plants, life cycles, natural enemies), which makes it very difficult to understand some of the patterns observed, for instance, the abundance vs. scarcity of some taxa. On the other hand, most entimines in Colombian collections are vegetation dwellers, whereas leaf litter, a highly diverse microhabitat for weevils (e.g. Anderson 2010), is essentially unexplored.

Most revisionary studies in northern South America have yielded numerous new species (Hustache 1938a; Howden 1976; Bordón 1991, 1997). With the notable exception of Howden (1976), not many taxonomic revisions of Neotropical entimines have included Colombian specimens; many of the specimens studied by Howden were collected directly by her and her colleagues in the early 1970s (Howden 1976). In the few recent revisions including Colombian material, examined specimens are housed in international collections and collected by early researchers in the times of “Nova Granada” (e.g. del Río and Lanteri 2007a, for *Melanocyphus*). This highlights a major issue for advancing the knowledge of Colombian entomofauna, in general: access to Colombian specimens by researchers is greatly limited. This fact, in turn, highlights other problems: (1) type material or even specimens correctly identified to species are extremely scarce (or non-existent) in national collections; (2) national experts (in Curculionidae), who would potentially be able to study type material abroad, are even more scarce, considering the high diversity of the group in the country (see Girón and Cardona-Duque 2018); (3) international experts essentially have no access to recent Colombian specimens, unless they visit Colombian national collections, which may not always be feasible; and, even if international researchers are able to visit, (4) national regulations make it very difficult to borrow specimens to take them out of the...
country for study. All of these issues combined, call for creating or strengthening international collaborations and adapting regulations to allow specimen exchange.

The good news is that digitisation efforts in libraries, and both in national and international biological collections, are contributing to improved access to specimens, data and expertise. Platforms, such as Biodiversity Heritage Library (https://www.biodiversitylibrary.org/), provide access to publications of original descriptions, museums are digitising their collections (entering data and imaging specimens, including types; for example, The Natural History Museum in London, (https://data.nhm.ac.uk/search/entomology) and SiB Colombia (https://sibcolombia.net/; https://colecciones.biodiversidad.co/) is providing access to specimen data from national collections. In addition, platforms, like iNaturalist (https://www.inaturalist.org/home), are continuously generating records for Colombian species. The greatest limitation to learning and understanding Colombian biodiversity is the availability of national experts who are able to study specimens both in national and international collections. Identifying entimines in Colombia is still a challenging task. Some of the specimens recorded here remain identified only to subfamily (Fig. 11) and are presented here in the hope that those can be revised and determined in the future, especially those from scarcely-explored regions.

Even though this work does not include all the entomological collections in the country and many remain to be revised, each collection revised here harbours a very unique diversity. It is evident that there is collecting bias towards the Andean region, with all other regions, but specially Amazon, Caribbean, Orinoco and the Islands (both Caribbean and Pacific), which are potentially as rich in diversity of species, poorly sampled sampled (see maps in Figs 2–11). The distribution of many species within the country is still to be determined: about one third of the species remain simply recorded from “Colombia” and about another third have been described from specimens collected in ‘Bogotá’ of the 1800s to early 1900s. Studying entimines in national collections and collecting them in poorly sampled regions will likely produce new records and taxa new to science all across the country.

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georeferencing all entimine records, revising and editing earlier versions of this manuscript and overall keeping me convinced that getting this compilation of information out in the world is valuable. Analía Lanteri has provided invaluable and critical feedback on several versions of this contribution over the years; thanks to her, the content of this document is much more accurate and comprehensive. The late Charles O’Brien and Nico Franz contributed their expertise, access to specimens and literature and encouragement at several points during the development of this work. Juan José Morrone also provided feedback and encouragement as a reviewer of the very first version of this manuscript, back in 2007. Alexander Ortiz facilitated the initial production of line-drawings. Ivon Babativa assisted databasing specimens at IAvH. My main motivation in producing this manuscript is to make information accessible and easy to find. It has taken me years to become familiar with literature about Colombian entimines and nowadays, I have the bibliographic resources, but living overseas, no access to Colombian specimens. I would like young researchers in Colombia to be able to quickly find information and to study entimine specimens right away! Many of the resources referenced here are available via The Biodiversity Heritage Library (https://www.biodiversitylibrary.org/), for which I’m also very grateful.

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**Supplementary material 1**

**DarwinCore-formatted list of species of Entiminae recorded from Colombia.**

This checklist is available via GBIF (Girón 2020, https://doi.org/10.15472/jdwfao)

Authors: Jennifer C. Girón

Data type: Species list

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Link: https://doi.org/10.3897/neotropical.15.59713.suppl1
Supplementary material 2

DarwinCore-formatted occurrence records for 749 entimine specimens deposited in Colombian biological collections
Authors: Jennifer C. Girón
Data type: Occurrence data
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Link: https://doi.org/10.3897/neotropical.15.59713.suppl2